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Patricia L. Gillis  
*University of Guelph*

James C. McGeer  
*Wilfrid Laurier University, jmcgeer@wlu.ca*

Gerald L. Mackie  
*University of Guelph*

Michael P. Wilkie  
*Wilfrid Laurier University, mwilkie@wlu.ca*

Josef D. Ackerman  
*University of Guelph*

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**Recommended Citation**

Gillis, Patricia L.; McGeer, James C.; Mackie, Gerald L.; Wilkie, Michael P.; and Ackerman, Josef D., "The Effect of Natural Dissolved Organic Carbon on the Acute Toxicity of Copper to Larval Freshwater Mussels (Glochidia)" (2010). *Biology Faculty Publications*. 33.  
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THE EFFECT OF NATURAL DISSOLVED ORGANIC CARBON ON THE ACUTE TOXICITY OF COPPER TO LARVAL FRESHWATER MUSSELS (GLOCHIDIA)

PATRICIA L. GILLIS,*† ‡ JAMES C. MCGEER,§ GERALD L. MACKIE, † MICHAEL P. WILKIE,§ and JOSEF D. ACKERMAN†

†Department of Integrative Biology, University of Guelph, Guelph, Ontario N1G 2W1, Canada
‡Aquatic Ecosystem Protection Research Division, Environment Canada, Burlington, Ontario L7R 4A6, Canada
§Department of Biology, Wilfrid Laurier University, Waterloo, Ontario N2L 3C5, Canada

(Submitted 28 September 2009; Returned for Revision 23 December 2009; Accepted 9 June 2010)

Abstract The present study examined the effect of dissolved organic carbon (DOC), both added and inherent, on Cu toxicity in glochidia, the larvae of freshwater mussels. Using incremental additions of natural DOC concentrate and reconstituted water, a series of acute copper toxicity tests were conducted. An increase in DOC from 0.7 to 4.4 mg C/L resulted in a fourfold increase (36 150 µg Cu/L) in the 24 h median effective concentration (EC50) and a significant linear relationship (r² 0.98, p 0.0008) between the DOC concentration and the Cu EC50 of Lampsilis siliquoidea glochidia. The ameliorating effect of added DOC on Cu toxicity was confirmed using a second mussel species, the endangered (in Canada) Lampsilis fasciola. The effect of inherent (i.e., not added) DOC on Cu toxicity was also assessed in eight natural waters (DOC 5–15 mg C/L). These experiments revealed a significant relationship between the EC50 and the concentration of inherent DOC (r² 0.79, p 0.0031) with EC50s ranging from 27 to 111 µg Cu/L. These laboratory tests have demonstrated that DOC provides glochidia with significant protection from acute Cu toxicity. The potential risk that Cu poses to mussel populations was assessed by comparing Cu and DOC concentrations from significant mussel habitats in Ontario to the EC50s. Although overall mean Cu concentration in the mussel’s habitat was well below the acutely toxic level given the concentration of DOC, episodic Cu releases in low DOC waters may be a concern for the recovery of endangered freshwater mussels. The results are examined in the context of current Cu water quality regulations including the U.S. Environmental Protection Agency’s (U.S. EPA) biotic ligand model. Environ. Toxicol. Chem. 2010;29:2519–2528, © 2010 SETAC

Keywords Copper toxicity Dissolved organic carbon Freshwater mussels Glochidia Biotic ligand model

INTRODUCTION

Freshwater mollusks are among the most endangered groups of organisms in North America, with nearly 70% of the species designated as either threatened, endangered, or in decline [1]. Their decline has been attributed to a number of factors, including habitat alteration, loss of fish hosts, invasive species, and exposure to environmental pollution [2–5]. Some studies [2,6] have suggested that environmental contamination may contribute to recruitment or reproductive failure in freshwater mussels. As with most organisms, the early life stages of freshwater mussels are the most sensitive to environmental pollution. The larvae, called glochidia, are obligate parasites on fish. Most species of freshwater mussels release their glochidia from the brooding chambers in the gills of the female into the water column, where on contact they will encyst upon a host. The amount of time that a given glochidium is in the water column and exposed to waterborne contaminants can vary significantly (seconds to weeks) depending on the specific reproductive strategies of the species (luring or broadcasting) and how long it takes to make contact with a host [7]. Although freshwater mussels are sensitive to a range of pollutants, they are particularly sensitive to Cu [6,8–11]. In fact, concerns have been expressed that water quality guidelines for Cu do not adequately protect the early life stages of freshwater mussels [12,13]. It is important that toxicity data for this sensitive early life stage be included when water quality regulations are developed, because contaminants that reduce glochidia success will put mussel populations at risk.

As with all metals, Cu bioavailability is controlled by the composition of the exposure water. Major ions, including Ca, Mg, and Na, compete with metal ions at the site of uptake [14,15]. Similarly, the presence of ligands, such as dissolved organic carbon (DOC), can bind with the metal ion and reduce its bioavailability [16]. Copper bioavailability is strongly influenced by DOC. Free Cu ions (Cu²⁺), the most bioavailable and thus toxic Cu species in solution [17], form complexes with the negatively charged natural organic matter, reducing the amount of free Cu in solution and thereby reducing the bioavailability of Cu [18]. In fact, Mantoura et al. [19] reported that greater than 90% of Cu in freshwater was complexed by humic materials. The effect of copper DOC complexation on the bioavailability and toxicity of Cu has been demonstrated in a range of aquatic organisms, from cladocerans to fish [20–25], but until recently information on the effect of DOC on Cu toxicity in freshwater mussels was relatively scarce. Wang et al. [26] evaluated the effect of natural DOC on acute Cu toxicity in juvenile freshwater mussels. They observed a significant linear increase in the Cu EC50 (7 12 fold) when exposure solutions were augmented with up to 10 mg/L of natural DOC. Markich et al. [27] investigated the effect of DOC (as fulvic acid) on the response of adult Hyridella depressa (Australian freshwater river mussel) to metals. They reported an 18 fold decrease in Cu sensitivity (as measured in terms of duration of valve opening) when the concentration of fulvic acid in the exposures was increased from 0 to 11 mg/L. Hanstén et al. [28] reported that additions of
Mussels were fed a commercial shellfish diet (Instant Algae) as well as the implications for populations of freshwater mussels. The present study, therefore, examines the effect of natural DOC, both added and inherent (i.e., not added) on the Cu toxicity of the larvae of two species of freshwater mussels. Specifically, the influence of DOC on acute Cu toxicity of glochidia was examined using a reconstituted soft water and incremental additions of a natural organic matter, and also using a selection of source waters with their inherent natural organic matter (DOC varied from 5 to 15 mg C/L). These data are discussed in relation to current water quality criteria as well as the implications for populations of freshwater mussels.

**MATERIALS AND METHODS**

**Mussel collection and laboratory care**

Gravid *Lampsilis fasciola* (Rafinesque 1820) and *Lampsilis siliquoidea* (Barnes 1823) (fatmucket) were collected from the Thames River, and the latter were also collected Cox Creek (Grand River Watershed), both in Ontario, Canada. *L. fasciola*, which has been designated as an endangered species in Canada by the Committee on the Status of Endangered Wildlife in Canada (COSEWIC), was collected under the Species at Risk Permit Number SECT 06 SCI 007. The mussels were main tained in the Aqualab facility at the University of Guelph, (Guelph, ON, Canada) in a flow through system using well water at 10 ± 2°C to prevent the release of glochidia. The mussels were fed a commercial shellfish diet (Instant Algae Shellfish Diet 1800®) which is a mixture of a *Nannochloropsis*, *Isochrysis*, *Pavlova*, *Thalassiosira*, and *Tetraselmis* algae at a rate of approximately 1.2 × 10⁹ algae cells per mussel per day.

Glochidia were collected by flushing the marsupia (brooding chambers) with water using a syringe. The viability of each mussel’s glochidia was assessed prior to use (viability procedure described below). For each exposure, glochidia were collected from a minimum of three gravid females that exhibited greater than 90% viability. Prior to initiating an exposure, the pooled glochidia sample was gradually (over a 2 h period) brought up to the exposure temperature of 21°C by dilutions with room temperature reconstituted water.

**Acute copper toxicity tests with glochidia**

Toxicity tests were modeled after the American Society for Testing and Materials method [29] for conducting toxicity tests with the early life stages of freshwater mussels. Briefly, glochidia were removed from gravid female mussels and exposed to various concentrations of a waterborne contaminant. After a given period of exposure (24 h), the viability of the exposed glochidia (i.e., ability to close their valves and clamp down on a fish’s gill for encystment) was assessed using a concentrated salt solution (NaCl 240 g/L). Glochidia viability was assessed in a subsample (100 to 200) of the approximately 1,000 glochidia exposed in each replicate. Viability was calculated using the following equation: Percent Viability = 100 × (Number of closed glochidia after addition of NaCl – Number of closed glochidia before addition of NaCl) / (Number of closed glochidia after addition of NaCl + Number of open glochidia after addition of NaCl). Results are expressed as EC50 values rather than median lethal concentration (LC50) values, but because they are obligatory parasites on fish, for practical purposes nonviable glochidia should be considered dead, because they would be unable to attach to a fish host and complete their life cycle.

An aqueous Cu stock solution made from certified American Chemical Society grade (Fisher Scientific) cupric sulfate (CuSO₄·5H₂O) was used to create the copper exposures. Copper spiked DOC exposure solutions (described below) were acclimated to the test temperature (21°C) before addition of glochidia. Exposures (100 ml per replicate) were conducted in 250 ml glass beakers, under a 16:8 h light:dark cycle in an environmental chamber maintained at 21 ± 2°C. As per the American Society for Testing and Materials method, exposure solutions were not renewed nor were the glochidia fed during the exposure. A composite water sample (10 ml) was collected from each treatment at the beginning of the exposure for determination of dissolved (i.e., filtered through an Acrodisc™ 0.45 μm in line syringe tip filter; PALL Life Sciences) Cu concentrations in the exposures. Water hardness, pH, dissolved oxygen, major ions (Ca, Mg, Na, K, SO₄, Cl), and alkalinity were also measured in water samples collected at the beginning each toxicity test. All glassware was washed with 10% nitric acid (Reagent Grade, Fisher Scientific) prior to use and all solutions were made with Millipore™ water.

**Effect of added DOC on copper toxicity**

Exposures with *L. siliquoidea* glochidia. The effect of DOC on the sensitivity of glochidia to Cu was assessed by conducting a series of exposures using reconstituted water and additions of concentrated natural organic matter. Dissolved natural organic matter was collected from Luther Marsh, near Guelph (ON, Canada) (43°57′N, 80°26′W). One hundred liters of marsh water (raw DOC = 78 mg C/L) was concentrated using a stainless steel portable reverse osmosis unit (Limnological Research) to produce a DOC concentrate of 747 mg C/L. In order to displace metals from the binding sites on the DOC and remove them from solution, the concentrate was treated with an H⁺ cation exchange resin (Amberlite, IR 118H) to a pH of 2.3. The concentrate was stored in the dark, at 4°C until it was used in the Cu toxicity tests, at which time the pH was adjusted to 7.0 using sodium hydroxide (Sigma, Canada).

Individual toxicity tests, referred to as exposures, were conducted at each of 0, 0.5, 1.0, 2.5, 3.75, 5.0, and 7.5 mg DOC/L (nominal) using the concentrated Luther Marsh DOC. Six Cu concentrations (treatments) were assessed in each exposure. Test solutions (water containing DOC and Cu) were created by adding the desired amount of DOC concentrate to 3 L of reconstituted soft water [30]. Then, to each of six 0.5 L aliquots of the DOC water the desired amount of Cu stock solution was added to create a range of Cu concentrations (nominal 0 to 200 μg Cu/L). The Cu spiked test solutions were held in the dark at 4°C for 48 h before initiation of the exposures to allow the Cu DOC mixture to equilibrate [31]. Acute toxicity tests were conducted as outlined above.

Single exposure with *L. fasciola* glochidia. To confirm the relationship between DOC concentration and the sensitivity of glochidia to Cu, we examined the effect of DOC on Cu toxicity
in a second, closely related mussel species, *L. fasciola*. Rather than conducting a series of separate exposures as was done with *L. siliquoidea*, a single exposure was conducted at a fixed, acutely toxic level of Cu while the concentration of DOC (i.e., treatments) was varied. A Cu exposure of 18 μg/L was chosen based on a 24 h EC50 of 17.6 μg Cu/L (95% confidence interval [CI] 14.2–22.6) reported by Gillis et al. [11] for this species. Test solutions were created by first adding 18 μg Cu/L (nominal) to 4 L of reconstituted soft water. Then, to each of seven 0.5 L aliquots of the Cu solution, the desired amount of DOC (nominal 0–7.5 mg C/L) was added to create the DOC treatments. The Cu spiked, DOC exposure solutions were held in the dark at 4°C for 48 h before initiation of the exposures. Acute toxicity tests (including water analysis) were conducted as outlined above. Although the 0 mg/L DOC exposure was the true control in this exposure it contained an acutely toxic concentration of Cu. Therefore, in order to assess the health of *L. fasciola* glochidia (i.e., true control survival) an additional treatment, consisting of only soft water (no Cu or added DOC), was conducted alongside the exposure.

### Effect of inherent DOC on copper toxicity

Water was collected from eight locations in southern Ontario, Canada (see Table 1 for locations) using acid washed 18 L high density polyethylene buckets. The sites were chosen to represent a gradient of inherent dissolved organic carbon (Table 1). No attempt was made to match other water characteristics (hardness, pH, etc.) between the field sites. In streams and rivers, the water sample was collected from just below the surface in an area where the water was visibly flowing. In lakes, the water was collected from an open water area at least 10 m offshore. Water samples were transported to the laboratory and filtered through a 0.45 μm filter (PALL Life Sciences) and then held in the dark at 4°C until used in exposures (maximum of two weeks). Copper was added (spiked) to each of the field collected waters to create a range of Cu concentrations (nominal, 0 to 200 μg Cu/L). The Cu spiked, field collected waters were held in the dark at 4°C for 48 h before initiation of the exposures. Toxicity tests were conducted in each of the eight Cu spiked waters with *L. siliquoidea* glochidia as outlined above.

### Copper analysis

The concentration of dissolved Cu in the test solutions were determined in a filtered (Acrodisk 0.45 μm in line syringe tip filter) water sample collected at initiation (t = 0) of the toxicity test. Copper was analyzed using a graphite furnace atomic absorption spectrophotometer (Varian 220FS SpectraAA, Varian Techtron). The detection limit for Cu was 0.02 μg/L. Method blanks (3) and Fisher Scientific calibration standards (every 20 samples) were included in every run. A maximum of 5% difference between duplicates was accepted.

### DOC analysis

The concentration of DOC was determined in water samples (10 ml, filtered using an Acrodisk 0.45 μm in line syringe tip filter) collected at the beginning of each exposure using a Shimadzu total organic carbon analyzer (model 5050A; Mandel Scientific). The detection limit for DOC was 0.1 mg/L. The concentration of total organic carbon in each sample was calculated automatically by subtracting inorganic carbon from total carbon [24].

### Statistical analysis

Percent glochidia survival is reported using means with standard errors (SE). The EC50s were determined by Probit Analysis (SPSS v. 11.0) using measured concentrations of dissolved copper (t = 0). The EC50s are presented with 95% CIs as EC50 (95% CI). The EC50s were considered to be significantly different when their 95% CI did not overlap [32]. If the 95% CI of two EC50s overlapped, then the Litchfield and Wilcoxon [33] method was applied to determine if they were significantly different. Survival curves were produced using the statistical analysis component (Regression Wizard) of Sigma Plot v. 10.0. Statistical differences (p < 0.05) in glochidia survival between treatments in the *L. fasciola* exposure were determined using analysis of variance (ANOVA) followed by Tukey’s test.

### Comparison of observed EC50 values to BLM predicted water quality criteria

To determine if the current water quality regulations for Cu in North America will protect larval freshwater mussels, the

### Table 1. The measured concentration (mg C/L) of dissolved organic carbon (DOC), and observed 24 h copper median effective concentrations (EC50) (95% confidence intervals) in exposures used to assess the effect of DOC on copper toxicity in *L. siliquoidea* glochidia; including a series of exposures employing reconstituted soft water augmented with natural DOC (Luther Marsh, ON) and a series of eight natural field collected waters (no added DOC) from Ontario, Canada. Field sites (water body, site name and location presented) were selected to cover a range of inherent DOC concentrations (5 to 15 mg C/L). The predictions of the U.S. Environmental Protection Agency’s copper biotic ligand model (BLM) version (2.2.3) [35] including the site specific final acute values (FAV) and criterion maximum concentrations (CMC) are given for each exposure.

<table>
<thead>
<tr>
<th>Water body</th>
<th>Site name</th>
<th>Latitude</th>
<th>Longitude</th>
<th>DOC (mg C/L)</th>
<th>Observed Cu EC50 (μg/L)</th>
<th>BLM predicted site specific FAV (μg/L)</th>
<th>BLM predicted site specific CMC (μg/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lake Huron</td>
<td>Eagle Harbour</td>
<td>44.18278</td>
<td>81.62917</td>
<td>5.0</td>
<td>30.9 (26.5, 36.3)</td>
<td>100.8</td>
<td>50.4</td>
</tr>
<tr>
<td>Lake Saint Clair</td>
<td>Pocket Bay</td>
<td>42.53064</td>
<td>82.61963</td>
<td>5.7</td>
<td>26.7 (20.3, 36.6)</td>
<td>85.7</td>
<td>42.9</td>
</tr>
<tr>
<td>Cold Water River</td>
<td>Cold Water</td>
<td>44.71250</td>
<td>79.64611</td>
<td>6.4</td>
<td>32.1 (21.1, 57.1)</td>
<td>135.4</td>
<td>67.7</td>
</tr>
<tr>
<td>Saugeen River</td>
<td>Hanover</td>
<td>44.16056</td>
<td>81.03194</td>
<td>6.9</td>
<td>71.4 (58.3, 86.5)</td>
<td>208.6</td>
<td>104.3</td>
</tr>
<tr>
<td>Thames River</td>
<td>London</td>
<td>42.97965</td>
<td>81.12756</td>
<td>9.4</td>
<td>90.9 (77.6, 106.7)</td>
<td>211.5</td>
<td>105.8</td>
</tr>
<tr>
<td>Grand River</td>
<td>Doon Heritage</td>
<td>43.40472</td>
<td>80.4333</td>
<td>11.3</td>
<td>104.4 (97.0, 111.0)</td>
<td>315.6</td>
<td>157.8</td>
</tr>
<tr>
<td>Sydenham River</td>
<td>Dawn Mills (Dresden)</td>
<td>42.58925</td>
<td>82.12872</td>
<td>14.7</td>
<td>110.9 (99.7, 124.6)</td>
<td>418.1</td>
<td>209.1</td>
</tr>
<tr>
<td>Thames River</td>
<td>Thamesford</td>
<td>43.05601</td>
<td>80.99274</td>
<td>14.8</td>
<td>97.6 (85.9, 99.1)</td>
<td>455.9</td>
<td>227.9</td>
</tr>
<tr>
<td>Soft water (SW)</td>
<td></td>
<td></td>
<td></td>
<td>0.7</td>
<td>36.1 (30.3, 44.0)</td>
<td>5.4</td>
<td>2.7</td>
</tr>
<tr>
<td>SW + 0.5 DOC</td>
<td></td>
<td></td>
<td></td>
<td>1.2</td>
<td>42.1 (34.0, 53.3)</td>
<td>6.9</td>
<td>3.4</td>
</tr>
<tr>
<td>SW + 1.0 DOC</td>
<td></td>
<td></td>
<td></td>
<td>2.0</td>
<td>63.7 (54.2, 78.6)</td>
<td>12.5</td>
<td>6.3</td>
</tr>
<tr>
<td>SW + 2.5 DOC</td>
<td></td>
<td></td>
<td></td>
<td>3.3</td>
<td>106.5 (85.4, 143.3)</td>
<td>32.7</td>
<td>16.4</td>
</tr>
<tr>
<td>SW + 3.75 DOC</td>
<td></td>
<td></td>
<td></td>
<td>4.4</td>
<td>150.2 (127.3, 187.0)</td>
<td>33.8</td>
<td>16.9</td>
</tr>
</tbody>
</table>
acute toxicity data produced in the present study were compared to Canadian Water Quality Guidelines (CWQG) and U.S. Ambient Water Quality Criteria (U.S. AWQC). Current CWQG are based on hardness of the receiving or testing water [34], whereas U.S. AWQC for Cu are derived using the U.S. EPA Cu biotic ligand model (BLM) [35] and a number of input parameters including temperature, pH, DOC, and major ion concentrations. The online version (2.2.3) of the BLM was used to derive exposure specific water quality criteria. The measured values for temperature, pH, DOC, Ca, Mg, Na, K, SO₄, Cl, and alkalinity from the exposures (see Table 2) were entered into the BLM program. The BLM output includes a final acute value (FAV) and a criterion maximum concentration (CMC) for each exposure; the BLM derived values were then compared to the observed 24 h Cu EC₅₀ values determined in this study.

Summary of water quality and mussel distribution data

Raw water quality data collected by the Provincial Water Quality Monitoring Network (PWQMN) were provided by the Ontario Ministry of the Environment (http://www.ene.gov.on.ca/en/publications/dataproducts). Copper (µg/L) and DOC (mg/L) data were provided for water samples taken between 1998 and 2008 covering a total of 134 sites across 10 Ontario conservation authorities (CAs). These 10 CAs, in particular the streams and rivers that flow through them, were selected after consultation with the chair of the Ontario Freshwater Mussel Recovery Team, (T.J. Morris, Canadian Department of Fisheries and Oceans, unpublished data) because they were considered to be significant mussel habitats. The conservation authorities selected for this summary along with their main mussel habitat (i.e., river) included: Ausable Bayfield CA (Ausable River), Grand River CA (Grand River), St. Clair Region CA (Sydenham River), Long Point Region CA (Lyn River), Maitland Valley CA (Maitland River), Quinte CA (Moira and Salmon Rivers), Saugeen Valley CA (Saugeen River), Niagara Peninsula CA (Welland River), and the Upper Thames River CA and Lower Thames Valley CA (Thames River) (Fig. 1). For the purposes of this summary, the water quality and mussel distribution data from the Upper and Lower Thames CAs were combined.

Individual data points (i.e., measured concentrations for either Cu or DOC) were averaged over time for each PWQMN site. For each site, an average 67 individual data points (i.e., chemical analysis conducted on a specific date) were used to calculate the “site average” for Cu and on average 17 individual data points were used to calculate the site average for DOC. Note that PWQMN did not initiate routine DOC analysis until 2001 or later and that DOC measurements were not taken at all locations (thus the lower number of DOC readings per site compared to Cu). These site averages were then averaged to determine an overall mean for each CA referred herein to as a CA mean. Site averages, rather than individual readings, were used to calculate each CA mean to prevent skewing of the CA mean due to differences in sampling frequency or extreme readings (e.g., unusually high concentrations). The CA range illustrates the lowest and highest individual measured values across the entire CA over the 10 years surveyed. Because the PWQMN copper detection limit was 0.8 µg/L, any readings below this level were set to 0 µg/L for purposes of calculating CA means.

Mussel distribution data were obtained from various sources ranging from individual watershed recovery plans to peer reviewed literature (Table 3). The total number of living mussel species found in each CA is, to the best of our knowledge, the most up to date assessment of mussel distribution in that CA using post 1996 [36] live animal survey data rather than the total number of species that have ever been found in that watershed (i.e., historic data). The number of endangered mussel species reported for each CA was obtained from a series of Department of Fisheries and Oceans (DFO) maps, which present the distribution of aquatic species at risk (fish and mussels) in each CA in Ontario (http://conservation.ontario.on.ca/projects/DFO.html).

RESULTS
Effect of added DOC on copper toxicity

*L. siliquoidea* exposures. The measured concentrations of DOC in the series of *L. siliquoidea* exposures were 0.7, 1.2, 2.0, 3.3, 4.4, 5.2, and 8.7 mg C/L. Control survival (24 h) was >80% in all exposures except in the two highest exposures (5.2 and 8.7 mg C/L), where it was slightly reduced (76 and 78%, respectively). Additions of natural organic matter (Luther Marsh) DOC significantly reduced the toxicity of Cu to *L. siliquoidea* glochidia (Fig. 2). In the control exposure without any added DOC (0.7 mg C/L), the 24 h EC₅₀ was 36.1 (30.3 44.0) µg Cu/L (Table 1). In comparison, the 4.4 mg C/L exposure produced an EC₅₀ of 150.2 (127.3 187.0) µg Cu/L and no EC₅₀ could be determined at the two highest DOC

Table 2. Summary of measured water chemistry including pH, dissolved organic carbon (DOC) (mg/L), and concentrations (mol/L) of calcium, magnesium, sodium, potassium, sulfate, chloride, and dissolved inorganic carbon (DIC) used to predict exposure specific water quality criteria using the online version (2.2.3) of the U.S. Environmental Protection Agency’s copper biotic ligand model (BLM) [35]*

<table>
<thead>
<tr>
<th>Site label</th>
<th>pH</th>
<th>DOC (mg/L)</th>
<th>Ca (mol/L)</th>
<th>Mg (mol/L)</th>
<th>Na (mol/L)</th>
<th>K (mol/L)</th>
<th>SO₄ (mol/L)</th>
<th>Cl (mol/L)</th>
<th>DIC (mol/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Soft water</td>
<td>7.45</td>
<td>0.7</td>
<td>2.01E 04</td>
<td>3.04E 04</td>
<td>6.96E 04</td>
<td>3.58E 05</td>
<td>5.22E 04</td>
<td>3.22E 05</td>
<td>6.89E 04</td>
</tr>
<tr>
<td>Soft water + 0.5 DOC</td>
<td>7.23</td>
<td>1.2</td>
<td>2.12E 04</td>
<td>3.18E 04</td>
<td>7.48E 04</td>
<td>3.73E 05</td>
<td>5.47E 04</td>
<td>3.44E 05</td>
<td>7.09E 04</td>
</tr>
<tr>
<td>Soft water + 1.0 DOC</td>
<td>7.29</td>
<td>2.0</td>
<td>2.25E 04</td>
<td>3.21E 04</td>
<td>7.83E 04</td>
<td>2.92E 05</td>
<td>5.42E 04</td>
<td>3.27E 05</td>
<td>7.57E 04</td>
</tr>
<tr>
<td>Soft water + 2.5 DOC</td>
<td>7.62</td>
<td>3.3</td>
<td>2.29E 04</td>
<td>3.60E 04</td>
<td>8.92E 04</td>
<td>4.69E 05</td>
<td>6.30E 04</td>
<td>3.86E 05</td>
<td>8.41E 04</td>
</tr>
<tr>
<td>Soft water + 3.75 DOC</td>
<td>7.43</td>
<td>4.4</td>
<td>2.04E 04</td>
<td>3.13E 04</td>
<td>8.05E 04</td>
<td>3.48E 05</td>
<td>5.43E 04</td>
<td>4.01E 05</td>
<td>7.78E 04</td>
</tr>
<tr>
<td>Lake Huron</td>
<td>8.30</td>
<td>5.0</td>
<td>6.51E 04</td>
<td>3.18E 04</td>
<td>1.80E 04</td>
<td>2.43E 05</td>
<td>1.71E 04</td>
<td>1.89E 04</td>
<td>1.68E 03</td>
</tr>
<tr>
<td>Lake Saint Clair</td>
<td>8.01</td>
<td>5.7</td>
<td>6.59E 04</td>
<td>3.36E 04</td>
<td>2.08E 04</td>
<td>2.56E 05</td>
<td>1.76E 04</td>
<td>2.20E 04</td>
<td>1.42E 03</td>
</tr>
<tr>
<td>Cold Water River</td>
<td>8.19</td>
<td>6.4</td>
<td>1.43E 03</td>
<td>6.50E 04</td>
<td>5.05E 04</td>
<td>3.89E 05</td>
<td>1.34E 04</td>
<td>5.27E 04</td>
<td>3.54E 03</td>
</tr>
<tr>
<td>Saugeen River</td>
<td>8.54</td>
<td>6.9</td>
<td>1.28E 03</td>
<td>1.13E 03</td>
<td>3.43E 04</td>
<td>2.92E 05</td>
<td>1.72E 04</td>
<td>3.95E 04</td>
<td>4.06E 03</td>
</tr>
<tr>
<td>Thames River (London)</td>
<td>8.18</td>
<td>9.4</td>
<td>1.82E 03</td>
<td>8.52E 04</td>
<td>1.58E 04</td>
<td>1.21E 04</td>
<td>6.48E 04</td>
<td>1.76E 03</td>
<td>3.55E 03</td>
</tr>
<tr>
<td>Grand River</td>
<td>8.42</td>
<td>11.3</td>
<td>1.26E 03</td>
<td>8.06E 04</td>
<td>1.45E 03</td>
<td>7.67E 05</td>
<td>3.49E 04</td>
<td>1.61E 03</td>
<td>3.11E 03</td>
</tr>
<tr>
<td>Sydenham River</td>
<td>8.47</td>
<td>14.7</td>
<td>2.02E 03</td>
<td>5.68E 04</td>
<td>6.52E 04</td>
<td>1.33E 04</td>
<td>4.66E 04</td>
<td>9.17E 04</td>
<td>4.31E 03</td>
</tr>
<tr>
<td>Thames River (Thamesford)</td>
<td>8.54</td>
<td>14.8</td>
<td>1.80E 03</td>
<td>9.63E 04</td>
<td>6.87E 04</td>
<td>1.03E 04</td>
<td>5.03E 04</td>
<td>8.49E 04</td>
<td>4.83E 03</td>
</tr>
</tbody>
</table>

*Temperature was 22°C in all exposures and the concentration of humic acid was set to 10%. Field site locations are given in Table 1.*
concentrations examined because there was no significant mortality at any of the Cu concentrations tested. The cause of the high mortality observed in the 40 mg Cu/L treatment of the 5.2 mg C/L DOC exposure (Fig. 2F) is unknown but an error in exposure solution preparation is suspected. A significant linear relationship \((r^2 = 0.98, p = 0.0008)\) existed between the concentration of DOC and the resulting Cu EC50 (Fig. 3).

L. fasciola exposure. The 24 h survival of L. fasciola glochidia in the soft water treatment without any added DOC or Cu was 97% (+0.7). The mean measured concentration of Cu in the exposures was 14.9 (+1.3) μg/L. The measured concentrations of DOC in the treatments were 1.5, 1.9, 2.4, 3.7, 4.7, 5.8, and 7.9 mg C/L. For the control copper treatment, i.e., the treatment that contained an acutely toxic amount of Cu but no added DOC, survival was 44.2% (+7.7) after 24 h of exposure. A

Table 3. Summary of copper (as total copper) and dissolved organic carbon (DOC) concentrations in significant (stream and river) mussel habitats in southern Ontario and the number of mussel species found in each habitat

<table>
<thead>
<tr>
<th>Conservation authority</th>
<th>CA mean DOC (mg C/L)</th>
<th>CA range DOC (^a) (mg C/L)</th>
<th>CA mean copper (^b) (μg/L)</th>
<th>CA range copper (^b) (μg/L)</th>
<th>Total no. mussel species [reference]</th>
<th>No. endangered mussel species (^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ausable Bayfield</td>
<td>5.8 (1.98), (n=3)</td>
<td>2.3 13.7</td>
<td>2.02 (0.67), (n=7)</td>
<td>0 9.6</td>
<td>23               [^d]</td>
<td>6</td>
</tr>
<tr>
<td>Grand River</td>
<td>6.0 (0.43), (n=13)</td>
<td>3.3 13.3</td>
<td>1.64 (0.64), (n=32)</td>
<td>0 20.1</td>
<td>25               [40]</td>
<td>9</td>
</tr>
<tr>
<td>St. Clair Region</td>
<td>6.7 (2.78), (n=3)</td>
<td>1.9 11.4</td>
<td>2.42 (1.22), (n=9)</td>
<td>0 12.8</td>
<td>34               [41]</td>
<td>11</td>
</tr>
<tr>
<td>Long Point Region</td>
<td>3.6 (NA(^a)), (n=1)</td>
<td>2.0 10.4</td>
<td>1.27 (NA), (n=1)</td>
<td>0 3.2</td>
<td>10               [^f]</td>
<td>5</td>
</tr>
<tr>
<td>Maitland Valley</td>
<td>5.6 (0.56), (n=5)</td>
<td>3.3 12.1</td>
<td>1.52 (0.42), (n=13)</td>
<td>0 33.6</td>
<td>9                [^b]</td>
<td>2</td>
</tr>
<tr>
<td>Quinte</td>
<td>NA</td>
<td>NA</td>
<td>0.97 (0.46), (n=17)</td>
<td>0 35.7</td>
<td>10               [36]</td>
<td>2</td>
</tr>
<tr>
<td>Saugeen Valley</td>
<td>4.7 (1.31), (n=3)</td>
<td>1.7 9.2</td>
<td>0.59 (0.28), (n=14)</td>
<td>0 4.3</td>
<td>8                [42]</td>
<td>2</td>
</tr>
<tr>
<td>Upper Thames River &amp; Lower Thames Valley</td>
<td>4.9 (0.75), (n=9)</td>
<td>2.9 11.7</td>
<td>2.08 (1.18), (n=38)</td>
<td>0 54.3</td>
<td>26 [43]</td>
<td>11</td>
</tr>
<tr>
<td>Niagara Peninsula</td>
<td>NA</td>
<td>NA</td>
<td>2.13 (1.18), (n=3)</td>
<td>0 8.6</td>
<td>10               [^f]</td>
<td>8</td>
</tr>
</tbody>
</table>

Water quality data (Cu and DOC) was provided by the Provincial Water Quality Monitoring Network (PWQMN) of the Ontario Ministry of the Environment ([39]; http://www.ene.gov.on.ca/en/publications/dataproducts). Mean water quality values and ranges (minimum maximum) are given for data collected from 1998 to 2008. Values reported as mean are the average of all site averages (repeated sampling at one site over time) for each conservation authority (CA). The number of individual site averages used to determine each CA mean or CA range is reported as \(n\), standard deviation is given in parentheses. Mussel distribution data are from various sources (see below).

\(^a\) PWQMN did not initiate routine DOC analysis until 2001 or later, and DOC measurements were not taken at all locations.

\(^b\) The PWQMN copper detection limit was 0.8 μg/L, therefore any copper concentration below this level has been reported as 0 μg/L.

\(^c\) Designated as such by the Committee on the Status of Endangered Wildlife in Canada.


\(^e\) Endangered species data are from the Canadian Department of Fisheries and Oceans (http://conservation.ontario.on.ca/projects/DFO.html).

\(^f\) NA data not available.

\(^g\) J.L. Metcalfe Smith, S.K. Staton, Environment Canada, Burlington, ON, Canada, unpublished data.

\(^h\) D.J. McGoldrick, J.L. Metcalfe Smith, Environment Canada, Burlington, ON, Canada, unpublished data.

\(^i\) Todd Morris, Department of Fisheries and Oceans, Burlington, ON, Canada, unpublished data.
significant increase in glochidia survival (compared to the control) was observed when concentrated Luther Marsh DOC was added to the Cu exposures. Glochidia survival in all DOC augmented treatments was more than 80%, with no significant difference observed between the treatments (1.9–7.9 mg C/L).

Effect of inherent DOC on copper toxicity

The concentration of naturally occurring (i.e., inherent) DOC in the field collected waters are given in Table 1. Control survival ranged from 86 to 95% in the eight field collected waters. The concentration of DOC had a significant effect on the sensitivity of *L. siliquoidea* glochidia to waterborne Cu exposure (Fig. 4). The 24 h EC50 values ranged from 30.9 µg Cu/L in water collected from Lake Huron (DOC, 5.0 mg C/L) to 110.9 µg Cu/L in water collected from the Sydenham River (DOC, 14.7 mg C/L). Regression analysis revealed a significant relationship ($r^2 = 0.79$, $p = 0.0031$) between the concentration of inherent DOC and the resulting Cu EC50 (Fig. 4).
The concentration of inherent DOC in natural waters had a significant linear relationship with the Cu EC50. The significance of DOC in natural waters was demonstrated by Nadella et al. [37] and De Schamphelaere et al. [22]. When DOC was increased from 0 to 5 mg C/L [26], this was nearly identical to the fourfold increase in the glochidia Cu EC50 reported in the present study when DOC was increased from <1 to 4.4 mg C/L using Luther Marsh DOC and reconstituted soft water. Additions of DOC have also been shown to reduce Cu toxicity in a number of aquatic species. In cladocerans, Giesy et al. [20] reported that the presence of organic matter reduced the accumulation of Cu in *Simopephalus serrulatus*, and De Schamphelaere and Janssen [21], as well as De Schamphelaere et al. [22] demonstrated the protective effect of natural DOC additions on the copper toxicity in *Daphnia magna*. Similarly, in fish (rainbow trout [24]; fathead minnow [18]) acute Cu toxicity has been shown to decrease when the concentration of DOC in the exposure water was increased. Nadella et al. [37] also recently demonstrated the ameliorating effect of added DOC on the Cu sensitivity of the embryos of marine mussels (*Mytilus trossolus*).

### Effect of inherent DOC on copper toxicity

The concentration of inherent DOC in natural waters had a significant effect on the sensitivity of glochidia to Cu. Copper sensitivity of *L. siliquoidea* glochidia exposed to Cu spiked water from Lake Huron (EC50 30.9 μg/L), the field collected water with the lowest concentration of DOC (5.0 mg C/L), was similar to the EC50 (36.1 μg Cu/L) in reconstituted soft water exposures; however, when acute copper toxicity was assessed in water collected from the Grand River (DOC 11.3 mg C/L), the resulting EC50 was nearly three times higher than that of the soft water exposure (EC50 104.4 μg Cu/L). A significant relationship between the concentration of inherent DOC and the toxicity of Cu was also demonstrated in the embryos of marine mussels [25]. Arnold [25] found that the Cu EC50 of *Mytilus galloprovincialis* was highly correlated ($r^2 = 0.71, p < 0.001$) to the concentration of inherent DOC in the water (54 natural marine waters tested).

It should be noted that dissolved organic carbon was not the only water chemistry parameter that varied across the natural waters employed in the present study; water hardness also ranged from 84 to 284 mg CaCO₃/L (Table 2 presents water composition of the field collected waters). Interestingly, there was a positive and significant correlation (Pearson correlation coefficient 0.81, $p = 0.015$) between water hardness and the concentration of inherent DOC in the natural waters used in the present study. The bioavailability of Cu is known to be influenced by a number of water chemistry parameters, including the concentrations of major cations (Ca, Mg, Na) [14,15]. Water hardness has been shown to decrease Cu sensitivity of glochidia in exposures using reconstituted waters. For example, *Epio blasma triquetra* glochidia exposed to Cu in moderately hard (166 mg CaCO₃/L) water had an EC50 twofold higher than those exposed in soft (40 mg CaCO₃/L) water [11]. Although the effect of water hardness is significant, increases in water hardness do not appear to affect Cu bioavailability as strongly as increases in DOC. Again, our earlier exposures with *E. triquetra* and Aldrich humic acid demonstrated that a small increase in the DOC also resulted in a doubling of the EC50 [11]. However, the presence of other toxicity modifying factors in our field collected water exposures explains the variability observed in the linear regression of DOC on EC50 for natural waters (Fig. 4) versus reconstituted waters (Fig. 2).
Comparison of added DOC and inherent DOC

On a per concentration basis, it appears that the Luther Marsh DOC, which was added to toxicity tests using reconstituted water, was much more effective at reducing the bioavailability of Cu than the natural DOC in the field collected waters. For example, exposures with 4.4 mg C/L of Luther Marsh DOC resulted in an EC50 of 150 μg Cu/L. In comparison, the EC50 values in Cu spiked natural waters with inherent DOC concentrations of 5 to 6 mg C/L all had EC50 values less than 40 μg Cu/L. A number of factors may have contributed to this difference including the source of the DOC. Schwartz et al. [24] compared natural organic matter (NOM) from a number of sources with respect to their ameliorating effect on metal toxicity. They demonstrated that the NOM from Luther Marsh, an optically dark NOM which is dominated by allochthonous (terrestrially derived) material, reduced Ag toxicity in fish to a greater degree than did other optically lighter NOMs. Typically, darker colored NOMs have more metal complexing abilities than the lighter autochthonous NOMs (i.e., those derived from aquatic photosynthesis) that are found in large lakes and oceans [23,24]. In fact, Luider et al. [23] reported that darker colored NOMs had about twofold greater Cu binding capacity compared to the lighter colored ones.

Although we did not isolate the natural organic matter from the field collected waters to quantify darkness (or composition) in the present study, based on earlier research it seems appropriate that the DOC from the large lakes and rivers we tested would be more autochthonous in nature (therefore less effective at binding Cu) and this may explain the relatively lower protective effects compared to Luther Marsh DOC. An alternate, or perhaps confounding, reason for the observed difference between the ameliorating capabilities of the Luther Marsh and other field collected DCs, may lie in the differences in the composition of the water used in the exposures. The reconstituted water used in the Luther Marsh exposures was much softer (40 mg CaCO3/L) than the natural waters used to assess the effect of inherent DOC (84–284 mg CaCO3/L). As discussed above, glochidia have been shown to be less sensitive to Cu when exposed in hard water compared to soft water [9,11]. The increased concentration of major ions (Ca2+, Mg2+) in hard water leads to increased competition between those ions and the copper ion (Cu2+) for binding sites on the organism [14,15]. In addition to competition for binding sites on the organism, there will also be competition among the various cations (Ca2+, Mg2+, Cu2+) for binding sites on the DOC molecules. Mantoura et al. [19] reported that Mg and Ca compete with Cu for binding sites on the humic acid molecule. Similarly, Iglesias et al. [38] found that the degree of Cu complexation with fulvic acid decreased as the concentration of Ca2+ in the exposure medium increased. This would indicate that DOC is more effective at reducing the bioavailability of Cu in soft water than in hard water. Therefore, we suggest that the enhanced protective ability of the Luther Marsh DOC in soft waters compared to the DOC from field collected waters is a result of both the increased metal binding capabilities of this allochthonous dominated DOC and the ion dilute soft water used in the exposures.

Copper sensitivity: water quality criteria and guidelines

Currently, Canadian Water Quality Guidelines for Cu are based on water hardness. For waters with hardness less than 120 mg CaCO3/L the guideline is 2 μg Cu/L [34]. For waters with hardness between 120 and 180 mg CaCO3/L, the guideline is 3 μg Cu/L [34]. These guidelines would be protective considering the EC50 values derived from the natural water exposures of the present study ranged from 27 to 111 μg Cu/L. Currently, the most scientifically advanced regulatory approach for Cu is the U.S. EPA Ambient Water Quality Criteria, which are derived on a site-specific basis using the biotic ligand model (BLM) [35]. Table 1 presents the observed EC50 values alongside the BLM predicted final acute values (FAVs) and criterion maximum concentrations (CMC) derived for the exposures in the present study. The CMC (or acute criteria) values (half the FAV) produced range between 43 and 228 μg Cu/L (Table 2). Typically, the actual water quality criteria, the criterion continuous concentration (CCC or chronic criteria), which is the maximum 4 d mean concentration that is allowed over a three year period, is derived by applying the acute to chronic ratio to the CMC. The application of the BLM (v. 2.2.3, in Cu WQC Calculation mode) produced predicted FAV values that for every natural field collected water were greater than the measured toxicity (Table 1). A number of explanations are possible. For example, it is possible that the natural waters came with a contaminant load that resulted in glochidia being sensitized to Cu. Alternatively, it is possible that DOC bound Cu is bioavailable to glochidia and contributes to toxicity. It is also possible that BLM predictions generally overestimate protective effects, particularly at elevated pH. All of the natural waters tested had pH values over 8.0 (range 8.1–8.6). In any event, the BLM was not able to predict Cu toxicity to glochidia in natural waters and set FAV values that were inappropriate. Whether the anomaly between measured and BLM predicted toxicity results from unique features of the organism or the model (or both) is unknown; however, it is deserving of further study.

Implications for native populations of mussels

The rivers and streams of the lower Great Lakes Basin contain the richest assemblage of freshwater mussel species in Canada [36]. Forty species of mussels are found in this area, including 11 federally endangered species (http://conservation.ontario.on.ca/projects/DFO.html). After surveying both historic (pre 1960) and more recent mussel distribution data of southern Ontario, Metcalfe Smith et al. [36] concluded that significant species losses and community changes in the freshwater mussel populations have occurred. They suggested that even by conservative estimates, somewhere between one sixth and one third of the mussel fauna in the Grand, Thames, and Sydenham Rivers have been lost. Although the decline of native mussels in North America has been attributed to a number of factors, including habitat alteration, loss of fish hosts, and the invasive zebra mussel, environmental pollution is also considered to have contributed to the decline of native freshwater mussels. However, until recently research in this area has been somewhat limited and thus the impact of waterborne contaminants on native mussel populations is still uncertain. In fact, all of the recovery strategies for freshwater mussel produced to date in Canada have indicated the “urgent need” to examine the threat that waterborne contaminants pose to the recovery of imperiled freshwater mussels. Therefore, to address this need, we examined the results of this laboratory based toxicity study in the context of the measured levels of Cu in the mussel’s natural habitat.

A summary of the copper (as total Cu) and DOC concentrations found in significant (stream and river) mussel habitats in Ontario over the past 10 years is presented in Table 3. Data were compiled by conservation authority. The number of...
Effect of DOC on Cu toxicity to glochidia

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mussel species found within each conservation authority area, as well as the number of endangered species, are also included. According to PWQMN data, the concentration of Cu found in mussel habitats in southern Ontario has ranged from less than detectable (<0.8 μg/L) to 54.3 μg/L over the past 10 years, and the mean Cu concentration (see Materials and Methods for explanation) across the CAs ranged from 0.6 to 2.4 μg Cu/L (Table 3). Although the Ontario Ministry of the Environment did not initiate routine DOC analysis until 2001, and DOC measurements were not taken at all locations, the data available indicate that the concentration of inherent DOC ranged from 1.9 to 13.7 mg C/L at these sites. The overall CA means for DOC spanned 3.6 to 6.7 mg C/L (Table 3). Therefore, according to our earlier [11] laboratory based acute toxicity tests using reconstituted soft water (no added DOC), the mean Cu concentration in the mussel habitats surveyed does not exceed the level found to be acutely toxic to even the most sensitive of the seven species tested (V. fabalis 24 h EC50, 6.9 μg Cu/L). This conclusion is supported by EC50 values derived from toxicity tests conducted in Cu spiked natural waters in the present study. For example, waters with similar levels of inherent DOC as the mean levels found in the CAs examined (4 to 7 mg C/L) produced EC50 values that ranged from 31 to 71 μg Cu/L. This is more than an order of magnitude above the mean level of Cu reported in the rivers examined. Even though the mean levels of DOC and Cu in the mussel habitats of southern Ontario indicate that exposure to acutely toxic levels of Cu is not the typical situation, the episodic pulses or spikes of Cu may be of concern, depending on the DOC concentration of the water at that time. For instance, in the Thames River, a habitat that supports 11 federally endangered species of mussels, Cu has been found to exceed 50 μg/L on occasion. With variation in DOC concentrations (for example, 2.9 11.7 mg C/L in the Thames River, PWQMN data) even short term spikes in Cu concentration could be harmful, especially if the spike occurs at a sensitive stage in the reproductive cycle, particularly during the time that glochidia are released into the water column (May October). The significant mussel habitats of southern Ontario are typically DOC rich and while there are many instances in nature in which excess nutrients and DOC can lead to a deterioration of water quality (i.e., eutrophication), the results of the present study indicate that the inherent DOC found in mussel habitats in southern Ontario should, on average, provide freshwater mussels with protection from acute Cu toxicity, although further study into the effects of episodic Cu release on endemic freshwater mussels would be beneficial.

Although the acute Cu sensitivity of glochidia has been assessed in a variety of mussel species [8,10,11], the diversity of the exposure water used in those studies was very limited. Typically, reconstituted waters have been used to determine an organism’s sensitivity to a contaminant and, although such standardized waters are useful for comparing sensitivities among species and among studies, they do not necessarily reflect the complexity of the water found in an organism’s natural habitat. Because freshwater mussels are found in a wide range of habitats from small urbanized streams to large oligotrophic lakes, it is important to understand their vulnerability to contaminants under such diverse conditions. By assessing the Cu sensitivity of glochidia in a range of natural waters we have demonstrated that the sensitivity of glochidia to Cu not only depends on the species [11], but also on the composition of the water, especially the concentration of DOC, in which they live.

CONCLUSION

In conclusion, the present study has demonstrated the significant protective effect that natural organic matter (as DOC) has on acute Cu toxicity in glochidia. The significant relationship between the concentration of inherent DOC and copper toxicity emphasizes that an understanding of the DOC level in the mussel’s natural habitat is necessary to assess the biotic ability of Cu and its potential threat to the sensitive early life stages of freshwater mussels.

Acknowledgement This work was supported by the World Wildlife Fund (Endangered Species Recovery Fund (WWF Canada)), the Copper Development Association, and the Species at Risk Spécies en Péril (Department of Fisheries and Oceans Canada). The authors thank R. Mitchell, K. McNichols, A. Schwalb, P. Ragaz, J. Klink, O. Birecua, and G. Nishihara for assistance in the laboratory and field, as well as A. Todd and S. Sunderani of the Ontario Ministry of the Environment for providing the raw water quality data (PWQMN), and T. Morris (DFO) for assistance selecting mussel habitats and making mussel distribution data available to us. O. Birecua was supported by an NSERC Metals in the Human Environment Strategic Network grant.

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