

Winter 2-2019

Identification of the alternative oxidase gene and its expression in the copepod *Tigriopus californicus*

Allison McDonald

Wilfrid Laurier University, amcdonald@wlu.ca

Carly E. Tward

Wilfrid Laurier University, gold8730@mylaurier.ca


Willie Cygelfarb

Wilfrid Laurier University, cyge3600@mylaurier.ca

Jaspreet Singh

Wilfrid Laurier University, jasp14@gmail.com

Follow this and additional works at: https://scholars.wlu.ca/biol_faculty

 Part of the [Biology Commons](#), [Cellular and Molecular Physiology Commons](#), and the [Comparative and Evolutionary Physiology Commons](#)

Recommended Citation

McDonald, Allison; Tward, Carly E.; Cygelfarb, Willie; and Singh, Jaspreet, "Identification of the alternative oxidase gene and its expression in the copepod *Tigriopus californicus*" (2019). *Biology Faculty Publications*. 82.

https://scholars.wlu.ca/biol_faculty/82

1 Title: Identification of the alternative oxidase gene and its expression in the copepod

2 *Tigriopus californicus*

3 Author names and affiliations: Carly E. Tward, Jaspreet Singh, Willie Cygelfarb and

4 Allison E. McDonald

5 Department of Biology, Wilfrid Laurier University, 75 University Ave. W., Waterloo,

6 Ontario, N2L 3C5, Canada

7 Corresponding author: Allison E. McDonald, amcdonald@wlu.ca; Department of

8 Biology, Wilfrid Laurier University, 75 University Ave. W., Waterloo, Ontario, N2L 3C5,

9 Canada

10

11

12

13

14

15

16

17

18

19

20

21 **Abstract**

22 In addition to the typical electron transport system (ETS) in animal mitochondria
23 responsible for oxidative phosphorylation, in some species there exists an alternative
24 oxidase (AOX) pathway capable of catalyzing the oxidation of ubiquinol and the
25 reduction of oxygen to water. The discovery of AOX in animals is recent and further
26 investigations into its expression, regulation, and physiological role have been
27 hampered by the lack of a tractable experimental model organism. Our recent DNA
28 database searches using bioinformatics revealed an AOX sequence in several marine
29 copepods including *Tigriopus californicus*. This species lives in tidepools along the west
30 coast of North America and is subject to a wide variety of daily environmental stresses.
31 Here we verify the presence of the AOX gene in *T. californicus* and the expression of
32 AOX mRNA and AOX protein in various life stages of the animal. We demonstrate that
33 levels of the AOX protein increase in *T. californicus* in response to cold and heat stress
34 compared to normal rearing temperature. We predict that a functional AOX pathway is
35 present in *T. californicus*, propose that this species will be a useful model organism for
36 the study of AOX in animals, and discuss future directions for animal AOX research.

37

38 Key Words: arthropod, electron transport system, environmental stress, mitochondria,
39 protein isolation, respiration, tide-pool

40

41

42 **Introduction**

43 The Alternative Oxidase (AOX) Pathway

44 Cellular respiration in animal cells takes place in the mitochondria via the
45 electron transport system (ETS). The ETS is comprised of multi-subunit complexes and
46 mobile electron carriers in the inner membrane of the mitochondria (Genova, 2014).
47 Four complexes, (I, II, III, and IV), and the mobile carriers ubiquinone/ubiquinol and
48 cytochrome c, shuttle electrons through the system to the final electron acceptor oxygen
49 (Genova, 2014). At the same time, several of these complexes move protons from the
50 matrix to the intermitochondrial membrane space and result in a proton motive force
51 (PMF) across the inner mitochondrial membrane (Genova, 2014). This PMF is used by
52 complex V (ATP synthase), and as protons flow back into the matrix through this
53 enzyme, ATP is synthesized through the process of oxidative phosphorylation (Genova,
54 2014).

55 Many animals also contain additional enzymes capable of putting electrons into
56 and/or removing electrons from the ETS (McDonald and Gospodaryov, 2018). One such
57 enzyme is the alternative oxidase (AOX), a terminal quinol oxidase that catalyzes the
58 oxidation of ubiquinol and the reduction of oxygen to water (McDonald and
59 Vanlerberghe, 2004; McDonald, 2008). In contrast to cytochrome c oxidase, which is
60 comprised of nuclear and mitochondria encoded subunits, the AOX is encoded by a
61 single nuclear gene (McDonald, 2008). The AOX protein is composed of four helices
62 and several conserved amino acid residues that are important for its enzymatic function
63 (McDonald, 2008). Use of the AOX pathway causes electrons to bypass complexes III

64 and IV and results in cyanide-resistant respiration (Rogov et al., 2014). As AOX is non-
65 proton pumping (in contrast to complexes I, III, and IV), it does not contribute directly to
66 the proton motive force (PMF), and the energy associated with electron transport is
67 dissipated as heat (McDonald, 2008). Therefore, AOX is viewed as energetically
68 inefficient.

69 AOX in Animals

70 AOX has been identified in several animal species due to the presence of AOX
71 DNA or mRNA sequences in public molecular databases (McDonald and Vanlerberghe,
72 2004; McDonald et al., 2009). Recent database searches have revealed the presence
73 of AOX DNA or mRNA in the phyla Ctenophora, Platyhelminthes, Arthropoda,
74 Tardigrada, Scalidophora, Brachiopoda, and Rotifera for the first time (McDonald and
75 Gospodaryov, 2018). Experimental evidence for the expression of AOX mRNA exists for
76 the sponge *Ephydatia muelleri*, and the molluscs *Anadara ovalis*, *Crassostrea gigas*,
77 *Crassostrea virginica*, and *Mercenaria mercenaria* (McDonald et al., 2009; Liu and Guo,
78 2017). Thus far, the only investigation of changes in AOX expression due to
79 environmental conditions has been the use of quantitative PCR (qPCR) to investigate
80 AOX transcript levels in the bivalves *Crassostrea gigas* and *Diplodon chilensis* which
81 increased in response to hypoxia and anoxia (Sussarellu et al., 2012; Yusseppone et
82 al., 2018).

83 The bulk of information about the AOX of animals exists at the level of genes or
84 transcripts. The only reports of naturally occurring AOX proteins present in animals are
85 from the brine shrimp *Artemia franciscana* and the Pacific oyster *Crassostrea gigas*;

86 AOX proteins in these species are ~35 kDa in size (Rodriguez-Armenta et al., 2018).
87 Knowledge of other animal AOX proteins is limited to studies that have taken advantage
88 of heterologous expression systems, therefore it is difficult to determine whether the
89 molecular masses of AOX reported reflect reality *in vivo*. For example, the expression of
90 the AOX from the sea squirt *Ciona intestinalis* produced a functional enzyme capable of
91 oxygen consumption of 42 kDa that was targeted to mitochondria in human kidney cells,
92 mice, and fruit flies (Hakkaart et al., 2006; El-Khoury et al., 2013; Kemppainen et al.,
93 2014).). Heterologous expression of the Pacific oyster AOX has also occurred in the
94 yeast *Saccharomyces cerevisiae* (Robertson et al., 2016). It is clear that there is a need
95 for an animal model that naturally expresses the AOX protein. The importance of finding
96 a suitable organism for the study of animal AOX is important as typically used model
97 systems (i.e. mouse and fruit fly) do not contain a naturally occurring AOX and therefore
98 cannot be studied (McDonald et al., 2009). No information exists regarding the post-
99 translational regulation of an animal AOX. In contrast to Complexes I-IV of the ETS,
100 there is no evidence that AOX exists as part of any respiratory supercomplex (Schertl
101 and Braun, 2014). The activation and inhibition kinetics of animal AOX proteins is also
102 unknown. Information on the characteristics of animal AOX proteins is needed, both due
103 to the desire for basic biological knowledge, and the realization that this knowledge is
104 required to assess the efficacy and safety of proposals to use AOX as a gene therapy
105 tool in humans for the treatment of mitochondrial diseases (El-Khoury et al., 2014). This
106 gap in our knowledge exists because to date animals known to contain AOX gene
107 sequences are often expensive to grow and culture in the lab, take up large amounts of
108 laboratory space, and do not have rapid life cycles which makes them impractical and

109 challenging for scientific study. In addition, research efforts have been hampered by the
110 existence of few genetic, molecular biology, and biochemical protocols and tools that
111 are available for use in these animals. It is clear that a worthy endeavor is to find a
112 useful AOX containing animal that is inexpensive to culture, has a rapid generation time,
113 takes up little space, and for which protocols and tool development is possible. In this
114 paper we assert that the intertidal copepod *Tigriopus californicus* meets all of these
115 criteria and can be used as a model system for the investigation of an animal AOX
116 protein.

117 *Tigriopus californicus*

118 *T. californicus* is an intertidal species of copepod found on the Pacific coast of
119 North America that inhabits rock pools (Burton and Lee, 1994). Due to *T. californicus*'
120 intertidal habitat, these animals are exposed to ever changing environmental stressors
121 including temperature, salinity, and oxygen levels (Burton and Lee, 1994) and
122 presumably have mechanisms in place to deal with such challenges. *T. californicus* has
123 a short generation time (~2-4 weeks depending on rearing temperature)(Hong and
124 Shurin, 2015), small space needs, and genetically divergent populations that can be
125 cross-bred in the laboratory (Burton & Feldman, 1981). A strong argument for using *T.*
126 *californicus* as a model organism in ecotoxicology and environmental genomics has
127 been made based on the above characteristics (Raisuddin *et al.*, 2007). In addition, a
128 protocol and tool exists for the suppression of gene transcription using RNA interference
129 in this species (Barreto *et al.*, 2015b).

130 *T. californicus* has been used to investigate the mechanisms of speciation and
131 incompatibilities in hybrid animals derived from mating different geographical
132 populations (Barreto *et al.*, 2015a). Hybrid incompatibilities have been demonstrated to
133 have a negative impact on oxidative phosphorylation and the interaction between the
134 nuclear and mitochondrial genome encoded processes (Barreto *et al.*, 2015a). This
135 incompatible hybridization causes mitochondrial dysfunction caused including inefficient
136 OXPHOS, a decrease in ATP biosynthesis, and increased ROS production (Barreto *et*
137 *al.*, 2015a). An investigation into the bioenergetics of *T. californicus* is therefore
138 warranted at the level of the ETS by examining complexes I-IV and AOX.

139 In this study we confirm the presence of the AOX gene, the expression of AOX
140 mRNA, and demonstrate that the AOX protein is present in the copepod *T. californicus*.
141 We believe this is the first step in establishing this copepod as a model species for
142 animal AOX research. By demonstrating that *T. californicus* is an effective model
143 species with experimental tools that are available, future research can examine the
144 physiological role of AOX in *T. californicus* and will explore questions about the
145 expression and regulation of AOX proteins in an animal system.

146 **Materials and Methods**

147 *In silico* identification of copepod AOX sequences

148 Molecular database searches using various animal AOX sequences in BLAST
149 (Basic Local Alignment Search Tool) using default settings at the National Center for
150 Biotechnology Information (NCBI) (<http://www.ncbi.nlm.nih.gov/>) (this was an iterative
151 process that used novel AOX sequences as they were discovered; full procedure as

152 described in McDonald et al., 2009) identified several putative copepod AOX
153 sequences. DNA sequences were translated into protein sequences using the ExPASy
154 Translate Tool (<http://web.expasy.org/translate/>).

155 Laboratory Culturing Conditions of *T. californicus*

156 Animals were obtained from J&L Aquatics Canada, Vancouver, British Columbia.
157 The animals arrived in a ~150mL bottle containing ~1000-2000 individuals at various life
158 stages. Animals used for DNA and RNA isolation were maintained in cultures of 20-200
159 individuals in 600 mL beakers containing 400mL of seawater (37.5 g/L) with a specific
160 gravity of 0.026 and grown at 21°C (room temperature) (Edmands and Deimler, 2004).
161 The salt water was made with Instant Ocean Salt and the specific gravity was tested
162 with the Instant Ocean Saltwater Aquarium Hydrometer (Instant Ocean, Blacksburg, VA,
163 USA). Each beaker was covered with the top plate of a petri dish to slow the rate of
164 evaporation. Beakers were placed in an incubator programmed to run on a 12 hour light
165 and 12 hour dark cycle. The animals were fed 4 drops of Reef Nutrition Phyto-Feast
166 Live Premium Phytoplankton (J&L Aquatics Canada, Vancouver, British Columbia)
167 every other day for the entire culturing period. Preliminary experiments demonstrated
168 that our animals completed their life cycle faster at 15.0°C, so a change in rearing
169 protocol occurred about 2 years into culturing *T. californicus* in the lab. Animals used for
170 protein isolations were cultured in 400mL beakers filled with 250mL of salt water (41.7 g
171 of Instant Ocean salt per 1L of MilliQ filtered water) at 15°C using a 12 hour light and 12
172 hour dark cycle. The copepods were fed once a week with 0.01 g of Tetramin fish food
173 (Nutrafin Basix) and 0.005 g of *Spirulina* powder (EarthRise) in 50mL of salt water
174 added to each habitat. The water in each habitat was changed weekly in order to

175 prevent the build-up of moulted exoskeletons and waste in the beakers. Animals used in
176 temperature experiments were acclimated to 15°C (normal growth temperature) for 2-4
177 weeks before being exposed to 6°C (cold) or 28°C (hot) for 24 hours (acute treatment)
178 or 1 week (chronic treatment) before collection for protein isolation.

179 DNA and RNA isolation and AOX Gene and Transcript analysis

180 Primer Design

181 Gene specific primers for the *T. californicus* 551 bp AOX cDNA fragment
182 (JW502496) identified in the NCBI database were designed using the Primer3 program
183 (<http://frodo.wi.mit.edu/primer3>). Custom primers were then synthesized (Invitrogen Life
184 Technologies, Carlsbad, CA, USA) for use in PCR and RT-PCR protocols (Table 1).
185 Three different forward and reverse primer sets were created in order to generate cDNA
186 products ranging from 200- 500bp; multiple primer sets were designed in case one or
187 more sets proved unsuccessful in producing a product.

188 Collection and preparation of copepods

189 In order to collect the copepods from the cultures for the isolation of DNA and RNA
190 the animals were strained out from the water using Miracloth (pore size: 22-25µm)
191 (Merck KGaA, Darmstadt, Germany). Once the water was completely drained from the
192 beaker, the copepods were washed several times with clean salt water and quickly
193 transferred to a pre-weighed Eppendorf tube, the mass of the animals was determined
194 (~200 mg), and the tube was frozen in liquid nitrogen. The frozen copepods were then
195 removed from the Eppendorf tube and ground into a fine powder using a previously
196 autoclaved and chilled pestle and mortar. The powdered sample was kept cold by
197 grinding under liquid nitrogen.

198 Isolation of nucleic acids and amplification parameters

199 For the extraction of DNA and RNA from *T. californicus* Trizol reagent proved
200 most effective (Invitrogen Life Technologies, Carlsbad, CA, USA) and the protocol was
201 conducted as per the manufacturer's instructions. Once the nucleic acids were
202 successfully isolated from *T. californicus* and quantified using spectrophotometry, PCR
203 and reverse transcriptase (RT)-PCR were conducted in order to amplify the targeted
204 AOX sequence. PCR was conducted with 5 µg of *T. californicus* DNA using 0.5 µL iTaq
205 DNA polymerase (Bio-Rad Laboratories Ltd., Mississauga, ON, CA), 2 µL of each 10
206 µM forward and reverse primer, 2 µL of 10x PCR buffer, 2 µL of dNTP mix, and 1.5 µL
207 of nuclease-free water in 20 µL total volume. The thermal cycler was set for 1 cycle of 2
208 minutes at 94°C for initial denaturation, 40 cycles consisting of: denaturation for 20
209 seconds at 94°C, annealing for 10 seconds at 58°C, and extension for 30 seconds at
210 68°C, followed by 1 cycle of final extension for 10 minutes at 68°C and the final soak
211 cycle at 4°C held indefinitely, until the sample was removed from the thermocycler (Bio-
212 Rad Laboratories Ltd., Mississauga, ON, CA).

213 RT-PCR was conducted using the Access RT-PCR Introductory System (Promega
214 Corporation, Madison, WI, USA) with 2 µg of total DNase treated RNA in 50 µL
215 reactions containing 10 µL AMV/Tfl 5x reaction buffer, 1 µL dNTP mix, 2 µL of each 10
216 µM forward and reverse primer, 2 µL of 25 mM MgSO₄, 1 µL AMV RTase, 1 µL Tfl DNA
217 polymerase, and 29.5 µL nuclease-free water. The thermal cycler was set for first strand
218 cDNA synthesis consisting of 45 minutes at 45°C for reverse transcription and 2
219 minutes at 94°C for AMV RT inactivation. This was followed by 40 cycles of
220 denaturation for 30 seconds at 94°C, annealing for 1 minute at 60°C, and extension for

221 2 minutes at 68°C, followed by 1 cycle of final extension for 7 minutes at 68°C and the
222 final soak cycle held at 4°C, until the sample was removed from the thermocycler (Bio-
223 Rad Laboratories Ltd., Mississauga, ON, CA).

224 The amplification products were separated using DNA gel electrophoresis using a
225 1.2% agarose gel containing 1mL 50x Tris-acetate-EDTA (TAE) buffer, 49 mL nuclease-
226 free water (dH₂O), and 0.60 g agarose powder run in a tank with 1x TAE running buffer.
227 5 µL 1 kb or 100 bp DNA ladder (GeneDirex, USA) was loaded in the first lane of the
228 gel, followed by 5-10 µL of each PCR or RT-PCR reaction combined with 6X DNA
229 loading buffer in subsequent lanes. Gels were run at 80-120V for 45 minutes and
230 stained with ethidium bromide in order to visualize the DNA under UV light (VersaDoc
231 400) (Bio-Rad Laboratories Ltd., Mississauga, ON, CA).

232 Isolation of Proteins from Copepods

233 For each protein extraction, approximately 0.05-0.10 g of copepods were isolated
234 from their habitats by filtration using a 100 µm filter (Fisherbrand) and placed in an
235 Eppendorf tube. Various combinations of different volumes of SDS-PAGE sample buffer
236 (350-500 µL) and 20 µL of β-mercaptoethanol solution were pipetted into each
237 Eppendorf tube. Each sample was sonicated (Omni International Inc.) for 10 seconds at
238 a frequency of 6 kHz and/ or homogenized to extract the proteins from the copepods.
239 The Precellys 24 lysis and homogenization machine (Bertin Technologies) was used to
240 perform the homogenization of copepod tissue. The homogenization processes
241 consisted of adding beads into each sample, loading the samples into the machine, and
242 3D bead-beating for 30 seconds at 5000rpm. These various combinations were used in

243 order to identify the optimal conditions for protein isolation from our laboratory grown
244 copepods.

245 Protein Analysis: Gel Electrophoresis

246 Protein samples were analyzed using reducing SDS-PAGE and Western blotting.
247 For gel electrophoresis, a 15 well mini-PROTEAN TGX Stain-Free gel (Bio-rad), was
248 loaded with 15 μ L of each copepod sample. In order to determine the molecular weight
249 of the proteins in our samples, two wells were loaded with 5 μ L of the Precision Plus
250 Protein WesternC Standards (Bio-rad). A yeast sample expressing the Pacific oyster
251 alternative oxidase (Robertson et al., 2006) served as a positive control in order to verify
252 the cross-reactivity of the AOX antibody with our copepod samples. A negative control
253 blot was run using proteins isolated from the algal food source to rule out the possibility
254 of an algal AOX being detected in our copepod samples. Following the loading of the
255 wells the gel was run at a constant voltage (200V) for 35 minutes. After the run was
256 complete, the gel was visualized using a VersaDoc (Bio-rad, USA) so an image could
257 be taken to visualize protein loading. For protein gels using our temperature experiment
258 samples, proteins in each sample were quantified using Quick Start Bradford Protein
259 Assay using bovine serum albumin standards and a spectrophotometre. Each sample
260 consisted of 20 μ g of protein added to each lane.

261 Protein Analysis: Western Blot

262 The Trans-Blot Turbo Transfer System (Bio-rad) was used to transfer the
263 proteins onto a nitrocellulose membrane in 3 minutes using Trans-Blot Transfer Medium
264 (Bio-rad). Following protein transfer, the membrane was placed in 5% dry milk in 10 mL

265 of 1X TBS-T on a shaking platform for 1 hour. The membrane was then subsequently
266 washed three times in TBS-T. 4µL of the primary AOX antibody (Plant AOX1/2, Cat. #
267 ABIN3197483, Agrisera Antibodies) was then added to 20 mL of 1X TBS-T for 1 hour
268 (dilution 1: 10,000). Following this, the membrane was again washed three times (15
269 minutes, 10 minutes, and 10 minutes) with TBS-T. The membrane was then incubated
270 in 2 µL secondary antibody, a goat anti rabbit IgG (Cat. # ABIN101988, Agrisera
271 Antibodies, dilution 1:25,000), and 2 µL of the Precision Protein StrepTactin HRP
272 Conjugate (Bio-rad) in 20mL of 1x TBS-T for 1 hour followed by three washes in 1X
273 TBS-T. It was subsequently placed in a mixture of luminol/enhancer and peroxide buffer
274 solution from the Immun Star Western C Chemiluminescent kit (Bio-rad) in a 1:1 ratio
275 for 5 minutes. The membrane was then placed in the VersaDoc, and the
276 chemiluminescent signal was detected and imaged.

277 **Results**

278 Identification of putative AOX sequences in copepods

279 A molecular database search using a BLAST (Basic Local Alignment Search
280 Tool) at the National Center for Biotechnology Information (NCBI) revealed the
281 presence of putative AOX sequences in 14 different copepod species (Table 2). The
282 copepod species are representatives of 3 different copepod orders and 12 different
283 families (Table 2). The predicted AOX proteins all contained at least 2 or more of the 4
284 iron-binding motifs (Table 2) containing conserved glutamate and histidine residues
285 required for the enzyme to be functional (McDonald et al., 2009). AOX DNA and mRNA

286 in copepods are present in multiple developmental stages, both sexes, and various
287 habitats around the world (Table 2).

288 In silico analyses of the *Tigriopus californicus* AOX sequence

289 A molecular database search revealed a putative *T. californicus* AOX DNA
290 sequence of 551 bp (Figure A.1). The predicted protein from *T. californicus* was a
291 partial sequence, however 3 out of the 4 iron binding motifs containing conserved
292 glutamate and histidine residues required for the enzyme to be functional (McDonald et
293 al., 2009) were present (Figure A.1). From previous studies, the C-terminal motif N-P-
294 [YF]-X-P-G-[KQE], was determined to be diagnostic for animal AOX proteins (McDonald
295 et al., 2009). *T. californicus* demonstrates a high level of motif conservation in the region
296 of the protein mentioned above (i.e. N- P-F-E-K-G-K) (Figure A.1).

297 Molecular analyses of the *T. californicus* AOX

298 Trizol reagent proved to be effective in isolating RNA from the copepods and
299 allowed us to proceed with RT-PCR. All three AOX primer sets yielded cDNA fragments
300 of the expected sizes (Figure 1). Proteins were successfully isolated from *T. californicus*
301 using 0.05 g of copepods and 350 or 500 μ L of SDS-PAGE sample buffer containing β -
302 mercaptoethanol and using various combinations of sonication and homogenization
303 (Figure A.2). The *T. californicus* AOX was recognized by a plant AOX antibody and was
304 ~ 50 kDa in size (Figure 2). The antibody failed to cross-react with the AOX protein
305 likely present in the algae provided to the copepods as food (Figure A.3). Previous
306 studies have demonstrated that algal AOXs contain features that differ from those of
307 other organisms (e.g. extended N-terminus and other insertion/deletions) (Nemanis et

308 al., 2013). We therefore expected that a plant AOX antibody would not cross-react with
309 the algal AOX.

310 Response of AOX Protein Levels to Temperature Treatments

311 Copepods were acclimated to 15°C for 2-4 weeks and then subjected to one of
312 three different temperatures (6, 15, or 28°C) for 24 hours or 1 week and then sampled.
313 Protein gels demonstrated that equal protein loading had occurred (Figure A.4 and A.5).
314 Western blots indicate that AOX protein levels are low under control conditions (24
315 hours at 15°C), but AOX levels are higher after 24 hours of exposure to 6°C and 28°C
316 (Figure 3). Similarly, AOX protein levels are low under control conditions (1 week at
317 15°C), but AOX levels are higher after 1 week of exposure to 6°C and 28°C (Figure 4).

318 **Discussion**

319 Discovery of AOX in Copepods and the Phylum Arthropoda

320 We have shown that AOX is in the genomes of multiple species of copepods
321 from around the world that inhabit a wide variety of ecological niches (Table 2.) The first
322 reports of AOX from arthropods were in the brine shrimp *Artemia franciscana* and
323 putative sequences from other members of the Chelicerata, Hexapoda, and Crustacea
324 (Rodriguez-Armenta et al., 2018; McDonald and Gospodaryov, 2018). It was previously
325 hypothesized that AOX was not present in arthropods due to a gene loss event
326 (McDonald et al., 2009), however, the above data refute this hypothesis.

327 Based on information gathered using bioinformatics tools, the copepod *T.*
328 *californicus* contains an AOX sequence (Figure A.1, Table 2). Our next goal was to

329 determine whether the AOX gene of *T. californicus* is expressed in the animal. RT-PCR
330 using RNA isolated from pooled developmental stages of the organism verified that
331 AOX mRNA is expressed in the copepod (Figure 1). This represents the first AOX
332 mRNA detected experimentally in any arthropod using gene specific primers for AOX.

333 In order to verify that the AOX sequence recovered from the arthropod *T.*
334 *californicus* was from this animal and not a contaminant (e.g. from a microbial symbiont
335 or pathogen) the animal's 551 bp cDNA sequence was translated to its predicted protein
336 sequence and was found to possess the C-terminal region N-P-F-E-K-G-K (Figure A.1).
337 This C-terminal motif is of particular interest because it is characteristic of only animal
338 AOXs and is different from the C-terminal regions observed in plants and fungal species
339 (McDonald et al., 2009).

340 We wished to confirm that AOX protein was detectable in *T. californicus*. Reducing
341 SDS-PAGE followed by Western blotting with a commercial plant AOX1/2 antibody
342 detected a protein of ~ 50 kDa in our copepod samples (Figure 2). Heterologously
343 expressed AOX proteins from *Ciona intestinalis* and *Crassostrea gigas* are 35-42 kDa in
344 size (Hakkaart et al., 2006; Robertson et al., 2016). The AOX protein of *Artemia*
345 *franciscana* is between 25 and 37 kDa on Western blots using the anti-AOX
346 *Sauromatum guttatum* monoclonal antibody (Rodriguez-Armenta et al., 2018). It is
347 expected that the food source (*Spirulina*) would contain AOX as it is an algae, however,
348 the AOX antibody did not recognize a protein in this sample (Figure A.3).

349 Finally, we wished to determine if AOX protein levels would change in response to
350 an environmental stressor. The copepods are normally reared at 15°C and have been

351 cultured at this temperature for several years and have likely adapted to this growth
352 temperature over many generations in our lab. We therefore selected 15°C as our
353 control temperature and exposed treatment groups to either 6°C (cold) or 28°C (hot) for
354 an acute time period of 24 hours or a chronic time period of 1 week prior to sampling the
355 animals. Animals grown at 15°C exhibited very low levels of detectable AOX protein
356 (Figures 3 and 4). In contrast, animals grown at 6°C or 28°C exhibited higher levels of
357 AOX protein after 24 hours of exposure (Figure 3) or 1 week of exposure (Figure 4).
358 Similar to what we report here, an increase in AOX protein levels in response to cold
359 stress has been reported in a variety of plants (McDonald, 2008). In addition, AOX1
360 protein levels sharply increase in response to heat stress in the green alga
361 *Chlamydomonas reinhardtii* (Zalutskaya et al., 2015). Unfortunately, no other studies
362 have examined the effects of biotic or abiotic stressors on AOX protein levels in any
363 animal that naturally possesses the enzyme. The only other investigation of a native
364 AOX from an animal is a Western blot confirming its presence in the brine shrimp
365 *Artemia franciscana* (Rodriguez-Armenta et al., 2018). Our finding is therefore the first
366 to demonstrate that the levels of a native AOX protein in an animal change in response
367 to an environmental stressor.

368 The Power of *T. californicus* as an Experimental System to Examine AOX

369 *T. californicus* represents an emerging model organism for research because of
370 its abundance, wide geographic distribution, ease of manipulation in laboratory settings,
371 and its ecological relevance due to its position in marine food webs (Raisuddin et al.,
372 2007). Lab grown cultures of *T. californicus* can be maintained for many generations
373 and are relatively inexpensive to house. We were able to isolate copepod DNA and

374 RNA using Trizol reagent and present a protocol here for successfully isolating proteins
375 from whole animals. Coupled with newer techniques such as RNA interference (RNAi)
376 (Barreto et al., 2015), it is clear that many experimental tools can be used to answer
377 questions about AOX in this organism.

378 Based on our results, we can begin to ask questions about the genes involved in
379 regulatory pathways that control transcript expression levels in response to
380 environmental changes, developmental stages, and gender. Analysis of AOX
381 expression levels can be conducted by subjecting *T. californicus* to a variety of
382 environmental stressors such as alterations in salinity and temperature (Lauritano et al.,
383 2012; Zhang et al., 2013) in order to observe patterns of AOX gene expression under
384 varying environmental conditions. The environmental stress of fluctuating salinity levels
385 is a key stressor to *T. californicus* and has been noted to affect the expression of a
386 variety of genes in the copepod (Burton and Lee, 1994; Van Aken et al., 2009). We
387 have demonstrated that AOX protein levels increase in *T. californicus* in response to
388 temperature stress, however, at this time it is not known why *T. californicus* has the
389 gene for and produces AOX protein. It is thought that the AOX pathway provides
390 metabolic flexibility and gives the organism the ability to survive under a multitude of
391 environmental stressors (Vanlerberghe, 2013). In addition, this system also represents
392 an opportunity to study the post-translational regulation of the AOX protein in *T.*
393 *californicus*.

394 Future Direction and Applications

395 Knowledge about animal AOX also has applications in human and animal
396 medicine. Comparative research may aid in the treatment of diseases caused by
397 parasitic protists, where AOX is a current target of drug design (May et al., 2017). AOX
398 research could lead to the development of anti-parasitic drugs that can be used to kill
399 parasitic copepods that live on the skin of economically valuable fish species. Humans
400 do not contain AOX, and the information gathered from our research may eventually
401 contribute to the treatment of mitochondrial dysfunctions and disorders in humans using
402 AOX gene therapy (El-Khoury et al., 2013; Fernandez-Ayala et al., 2009; Hakkaart et
403 al., 2006; Kemppainen et al., 2014). Heterologous expression systems provide a
404 potential route of expression to rescue electron flow and test hypotheses in order to
405 mitigate the deleterious complications involved with respiratory chain dysfunctions (such
406 as Parkinson's, diabetes mellitus, and deafness) (Hakkaart et al, 2006; Kemppainen et
407 al., 2014), but this requires extensive knowledge about the regulation of AOX gene
408 expression and the post-translational regulation of the AOX protein.

409 We believe that *T. californicus* is an excellent model system in which to study
410 AOX. These animals can be used to study the physiological role of AOX and to test
411 functional hypotheses about the enzyme. This will lead to an increase in our knowledge
412 of AOX in general and contribute to the assessment its future applications in healthcare
413 and aquaculture.

414 **Acknowledgements**

415 This work was supported by the Natural Science and Engineering Research
416 Council of Canada (NSERC), the Canada Foundation for Innovation Leaders

417 Opportunity Fund, the Ontario Research Fund, and the Faculty of Science Students'
418 Association (FOSSA) of Wilfrid Laurier University.

419 The authors report no conflict of interest.

420 **Figure Captions**

421 Figure 1. Reverse-transcriptase PCR products using *T. californicus* AOX gene specific
422 primers with DNase treated RNA. Lane 1, 5 μ L 100bp DNA ladder; lane 2, 10 μ L AOX1
423 primer set product (232bp product size); lane 3, 10 μ L AOX2 primer set product (344bp
424 product size); lane 4, 10 μ L AOX3 primer set product (503bp product size); lane 5, 10
425 μ L positive control from kit (323bp product size).

426 Figure 2. Detection of *T. californicus* AOX protein by reducing SDS-PAGE and Western
427 blotting using a plant AOX antibody. All *T. californicus* samples per well are 14 μ L of
428 protein samples derived from 0.05 g of copepods of mixed developmental stages
429 subjected to either sonication (S), homogenization (H), or a combination of the two (H &
430 S). The protein sample buffer volume used (in microliters) is indicated by the number
431 below the isolation method. Ladder lanes contain 5 μ L of the Precision Plus Protein
432 Western C Standards. The positive control is 14 μ L of a protein sample from isolated
433 mitochondria from *Saccharomyces cerevisiae* overexpressing the *Crassostrea gigas*
434 AOX.

435 Figure 3. Detection of *T. californicus* AOX protein by reducing SDS-PAGE and Western
436 blotting using a plant AOX antibody under 24 hours exposure to different temperatures.
437 All *T. californicus* samples per well are 20 μ g of protein sample derived from 0.05-0.10 g
438 of copepods of mixed developmental stages. Ladder lanes contain 5 μ L of the Precision

439 Plus Protein Western C Standards. Temperature treatments are indicated above each
440 sample well.

441 Figure 4. Detection of *T. californicus* AOX protein by reducing SDS-PAGE and Western
442 blotting using a plant AOX antibody under 1 week exposure to different temperatures.
443 All *T. californicus* samples per well are 20 µg of protein sample derived from 0.05-0.10 g
444 of copepods of mixed developmental stages. Ladder lanes contain 5µL of the Precision
445 Plus Protein Western C Standards. Temperature treatments are indicated above each
446 sample well.

447 Figure A1. cDNA and predicted protein sequences of the AOX from *Tigriopus*
448 *californicus*. The highlighted region contains the C-terminal motif that is highly
449 conserved in animal AOX proteins.

450 Figure A2. Protein samples run on reducing SDS-PAGE. Ladder lanes contain 5µL of
451 the Precision Plus Protein Western C Standards. The positive control is 14 µL of a
452 protein sample from isolated mitochondria from *Saccharomyces cerevisiae*
453 overexpressing the *Crassostrea gigas* AOX. All *T. californicus* samples per well are
454 14µL of protein samples derived from 0.05 g of copepods of mixed developmental
455 stages subjected to either sonication (S), homogenization (H), or a combination of the
456 two (H & S). The protein sample buffer volume used (in microliters) is indicated by the
457 number below the isolation method.

458 Figure A3. Protein samples run on reducing SDS-PAGE and Western blotted using a
459 plant AOX antibody. The ladder lane contains 5µL of the Precision Plus Protein Western
460 C Standards. The algae lane contains a sample derived from the *Spirulina* used to feed

461 the copepods. This sample was sonicated in the same manner as the copepod samples
462 and 14µL were loaded into the well.

463 Figure A4. Protein samples run on reducing SDS-PAGE. Ladder lanes contain 5µL of
464 the Precision Plus Protein Western C Standards. All *T. californicus* samples per well are
465 20 µg of protein samples derived from 0.05-0.010 g of copepods of mixed
466 developmental stages subjected to 24 hours at the temperatures indicated above the
467 wells of the gel.

468 Figure A5. Protein samples run on reducing SDS-PAGE. Ladder lanes contain 5µL of
469 the Precision Plus Protein Western C Standards. All *T. californicus* samples per well are
470 20 µg of protein samples derived from 0.05-0.010 g of copepods of mixed
471 developmental stages subjected to 1 week at the temperatures indicated above the
472 wells of the gel.

473

474

475

476

477

478

479

480

481

482

483

484

485

486

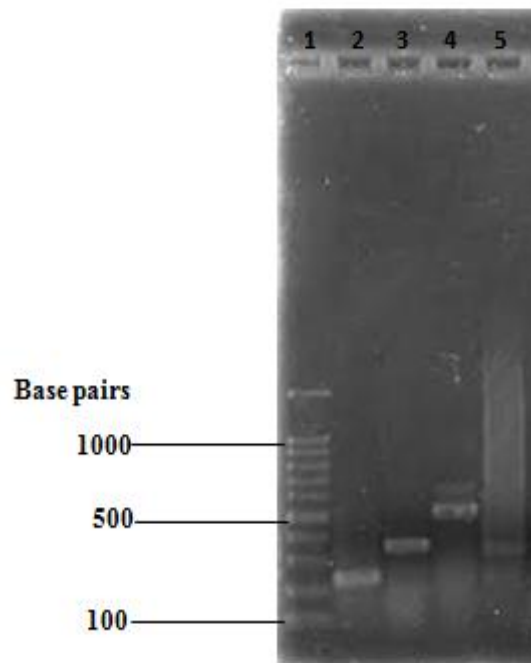
487

488

489

490

491



492

493

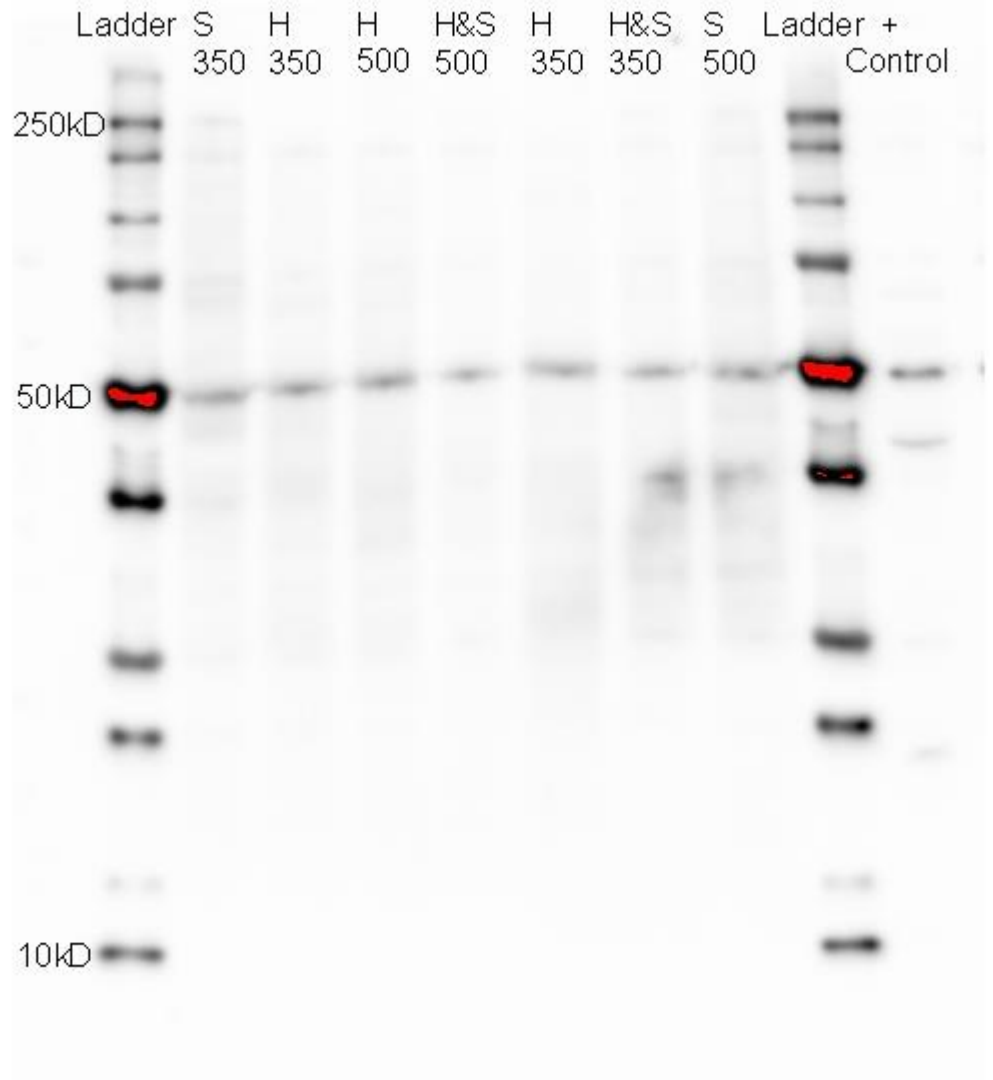
494

495

496

497 Figure 1.

498



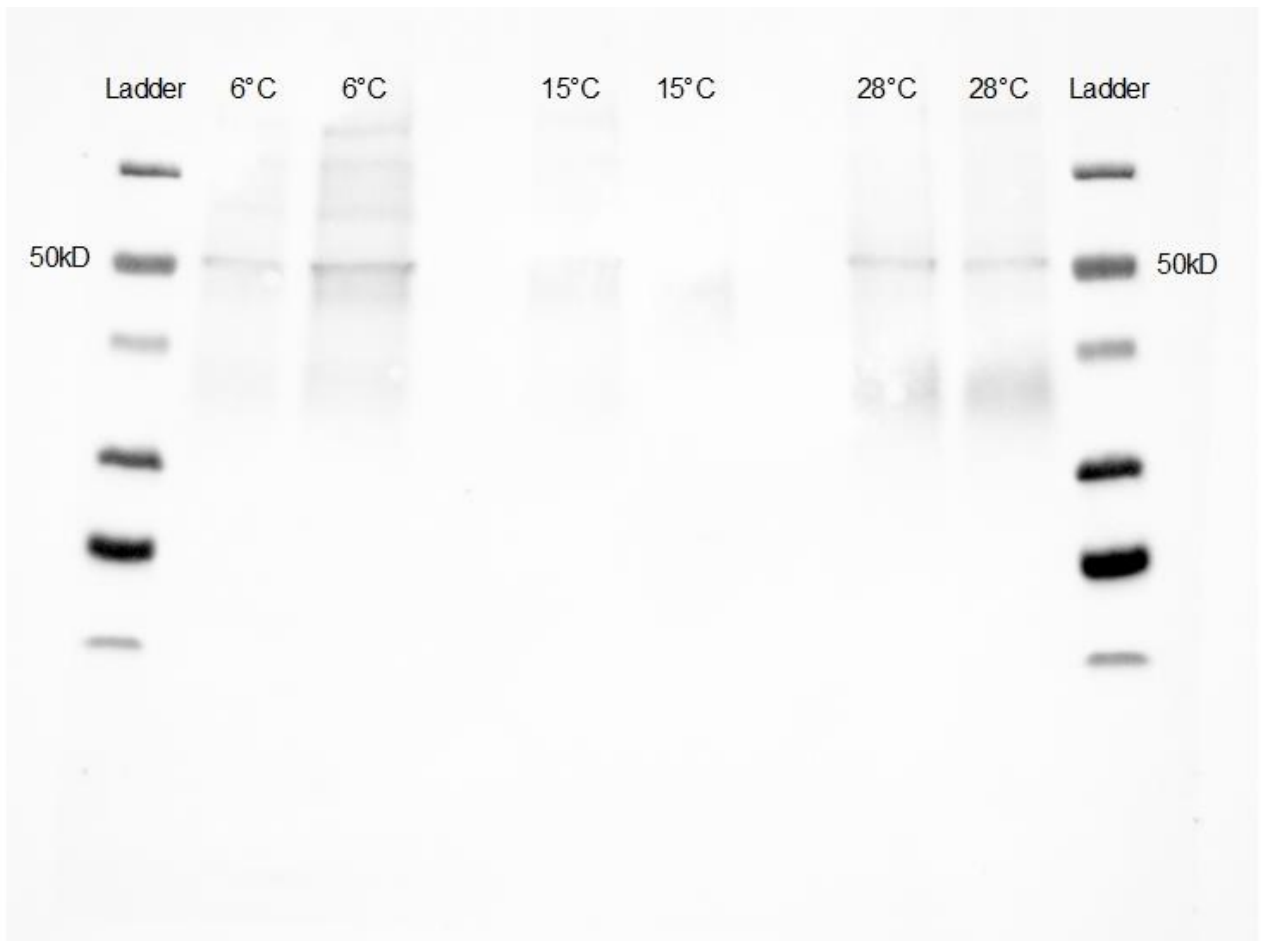
499

500

501

502 Figure 2.

503



504

505

506

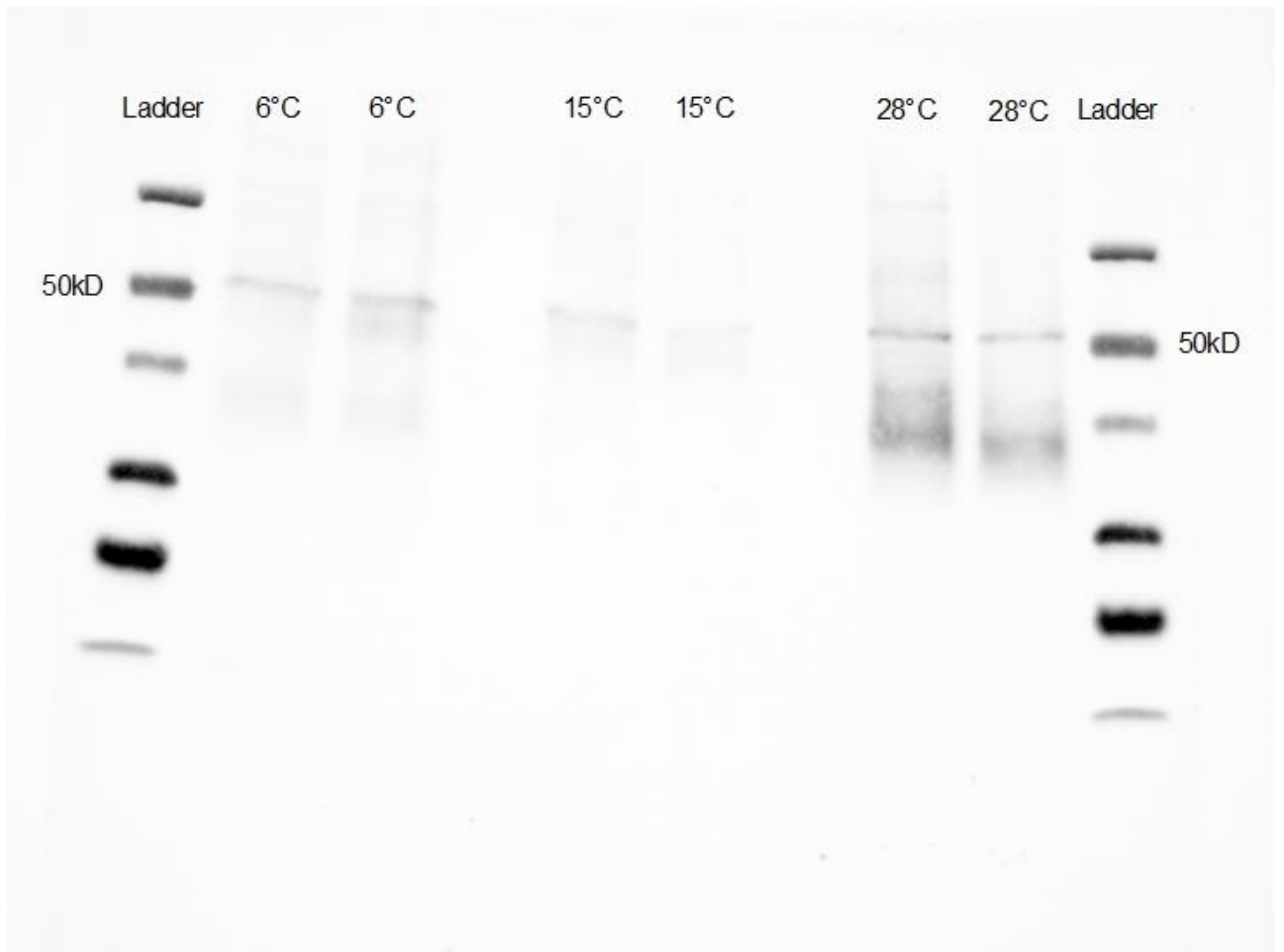
507

508

509

510 Figure 3.

511



512

513

514

515

516

517

518 Figure 4.

519

520

521

522

523 *Tigriopus californicus* 551bp cDNA sequence

524 GAATGCACGTGATCATGGATGGATCCACACTCTCCTGGAAGAAGCGGAGAATGAA
525 AGGATGCATCTGATGACCTTCATGCGACTCCGAAGACCCGGGCCCATTTTCCGAG
526 GCACCGTGATCCTGACTCAATGGCTGTTACATTTACGTTCTCATTGCTTACATAC
527 TGTCGCCCAATTTTTGCCACAGATTTGTTGGGTATTTGGAAGAGCAAGCAGTGGTC
528 ACTTACACTCACATCCTGGAAGAAATCGACGCAGGACGATTGCCCATGTGGAAGA
529 CCTTGCCAGCTCCGGAATTGGCCATCAAGTATTGGAGATTGCCCGAAGACGCCAA
530 GATGCGGGAAGTCATTTTGGCAATCCGAGCCGATGAAGCTCATCATCGGCTTGTG
531 AATCACACCCTTGGATCGATGGACCTCAAATCAGACAATCCTTTTGAGAAAGGGAA
532 ATAACTTTTGTCTTCGGCTCAGATACAAATTTAATGGTCAATAACAAGTATTCCATG
533 GTAAAAACGAACGGAGGTTGAGAAATATAAATAAATTGTCGAGTCTTTTC

534

535 *Tigriopus californicus* predicted protein sequence

536 NERMHLMTFMRLRRPGPIFRGTVILTQWLFTFTFSFAYILSPNFCHRFGVYLEEQAVVT
537 YTHILEEIDAGRLPMWKTLPAPELAIKYWRLPEDAKMREVILAIRADEAHHRLVNHTLGS
538 MDLKDNPFKEKGK

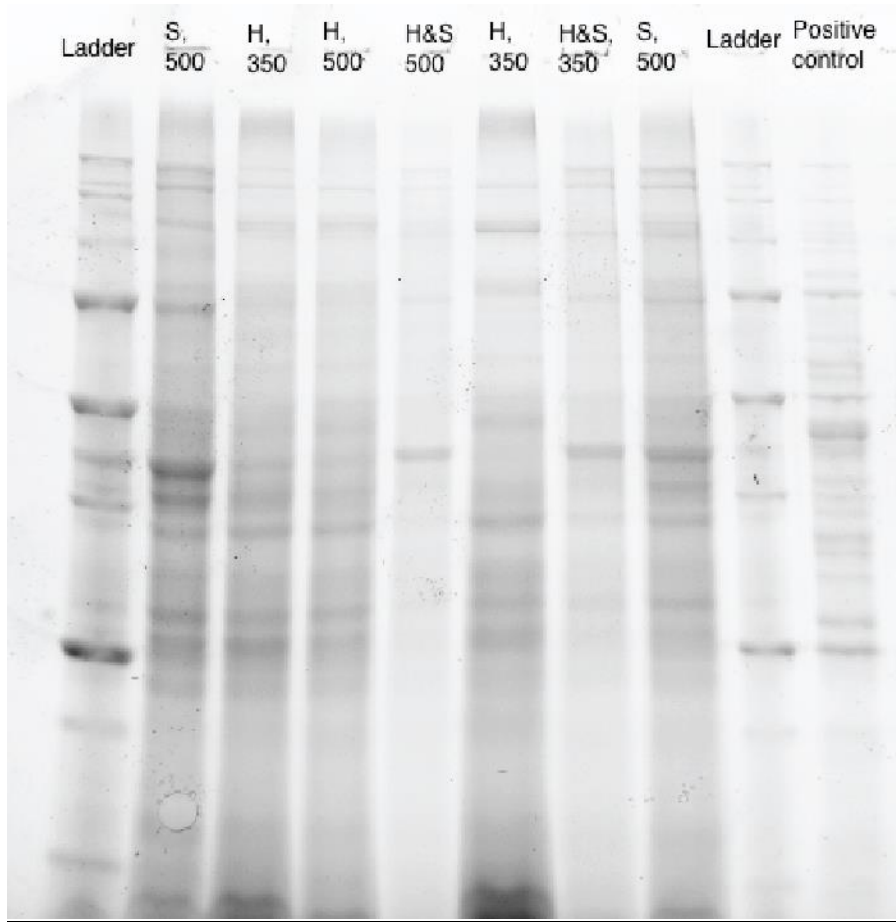
539

540 Figure A1.

541

542

543



544

545

546

547

548 Figure A2.

549

550

551

552

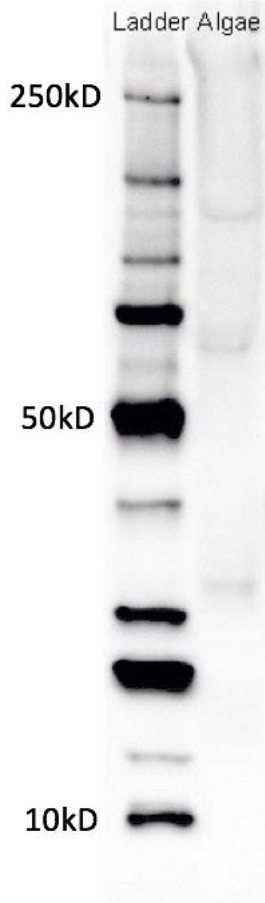
553

554

555

556

557



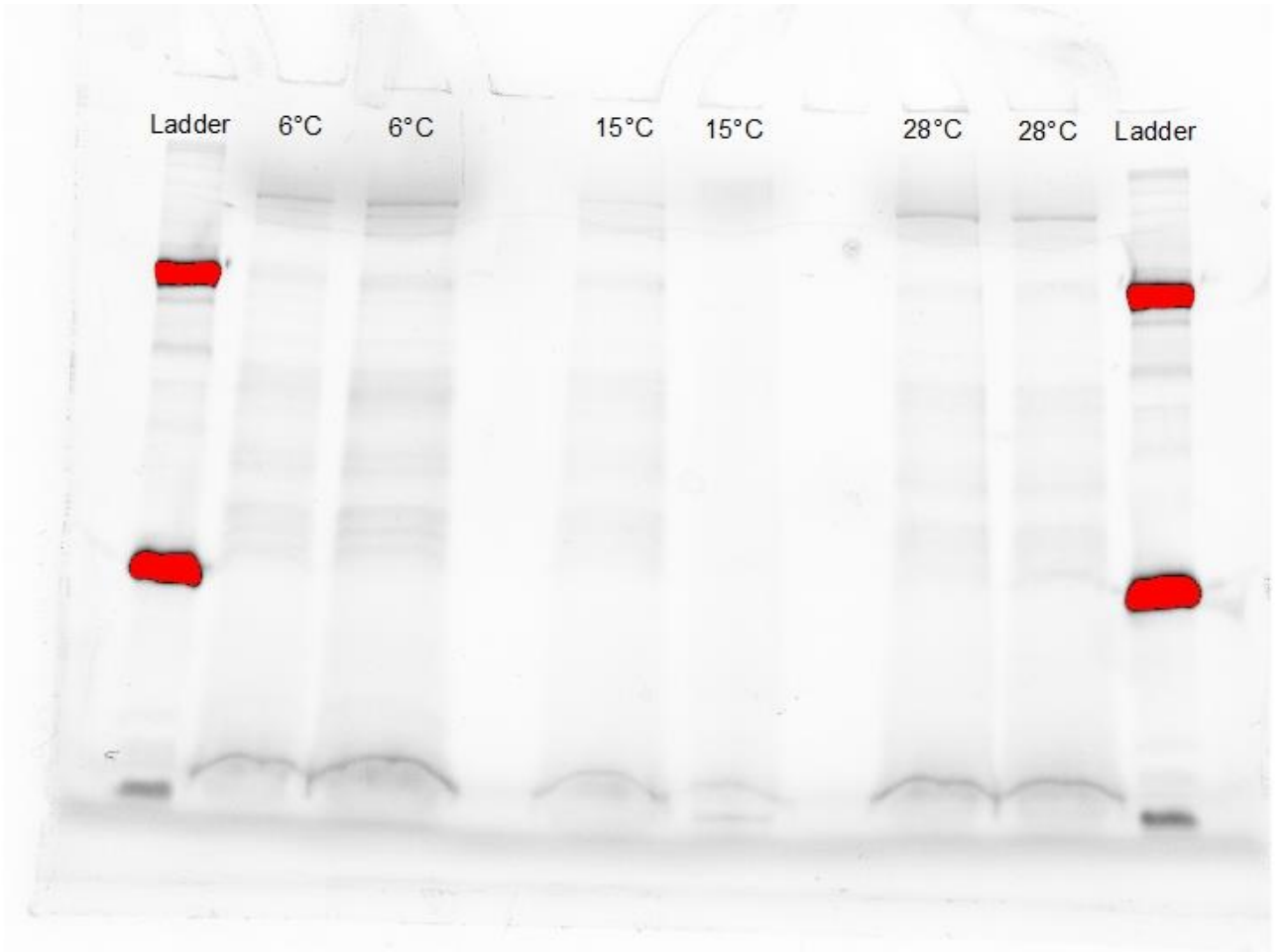
558

559

560

561

562 Figure A3.



563

564

565

566

567

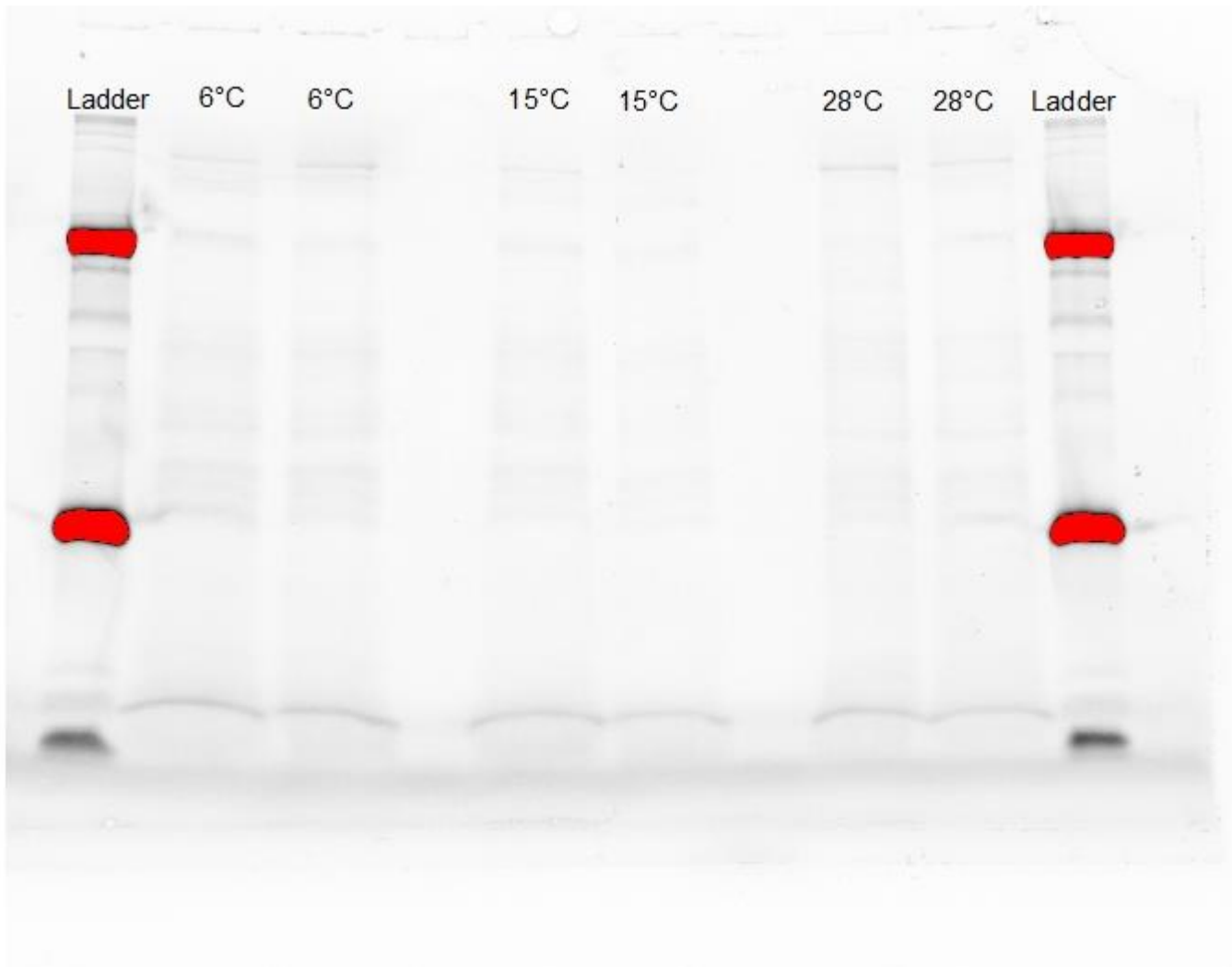
568

569

570 Figure A4.

571

572



573

574

575

576

577

578

579 Figure A5.

580

581

582

583 Table 1. Gene-specific primers designed for *Tigriopus californicus* AOX amplification.

584

| Primer Name | Forward Primer Sequence (5' → 3') | Reverse Primer Sequence (5' → 3') | Expected cDNA Product Size (bp) |
|--------------------|--|--|--|
| AOX1 | CCTGACTCAATGGCTGTTCA | CAAAATGACTTCCC GCATCT | 232 |
| AOX2 | TGCCACAGATTTGTTGGGTA | TTCTCAACCTCCGTTTCGTTT | 344 |
| AOX3 | GGATCCACACTCTCCTGGAA | CTCAACCTCCGTTTCGTTTTT | 503 |

585

586

587

588

589

590

591

592

593

594

595

596

597

598

599

600 Table 2. Putative AOX sequences in copepods recovered using bioinformatics searches
 601 of public databases.

| Order | Family | Species | Representative Accession Number | Iron-binding sites present in translated AOX protein | Information on Source Material |
|------------|---------------|------------------------------|---------------------------------|--|--|
| Calanoida | Acartiidae | <i>Acartia tonsa</i> | HAGX01001020 | All 4 | All developmental stages |
| | Calanidae | <i>Calanus finmarchicus</i> | GAXK01135432 | All 4 | C5 copepodite, Trondheim, Norway |
| | | <i>Calanus glacialis</i> | HACJ01021827 | IBS 1, 2, & 3 | Whole organism, Norway |
| | | <i>Neocalanus flemingeri</i> | GRUD01065088 | All 4 | Adult female |
| | Caligidae | <i>Caligus rogercresseyi</i> | GAZX01011857 | IBS 1, 2, & 3 | Mixed developmental stages and sexes |
| | Metridinidae | <i>Pleuromamma xiphias</i> | GFCI01346245 | All 4 | Whole adults, both sexes, BATS, Atlantic Ocean |
| | Pontellidae | <i>Labidocera madurae</i> | GFWO01190644 | All 4 | |
| | Temoridae | <i>Eurytemora affinis</i> | GBGO01037152 | All 4 | Whole females |
| Cyclopoida | Cyclopettidae | <i>Paracyclopina nana</i> | GCJT01014574 | All 4 | Whole animals, Gangneung, South Korea |
| | Cyclopidae | <i>Eucyclops serrulatus</i> | GARW01011119 | All 4 | |
| | Oithonidae | <i>Oithona nana</i> | FTRT01002078 | IBS 1 & 2 | |

| | | | | | |
|---------------|---------------|--------------------------------|--------------|-------|---------------------------------------|
| Harpacticoida | Harpacticidae | <i>Tigriopus californicus</i> | JW502496 | All 4 | San Diego population |
| | Harpacticidae | <i>Tigriopus japonicas</i> | GCHA01002206 | All 4 | Whole animals, Gangneung, South Korea |
| | Harpacticidae | <i>Tigriopus sp. 1 SL-2012</i> | GDFW01016749 | All 4 | Tidal pool, Maxwell Bay, Antarctica |

602

603 ¹ Trace sequence archive database at NCBI

604 ² Nucleotide database at NCBI

605 References

606 Barreto, F.S., Pereira, R.J., Burton, R.S., 2015a. Hybrid Dysfunction and Physiological
607 Compensation in Gene Expression. *Molecular Biology and Evolution*. 32, 613-622.

608

609 Barreto, F.S., Schoville, S.D., Burton, R.S., 2015b. Reverse genetics in the tide pool:
610 knock-down of target gene expression via RNA interference in the copepod *Tigriopus*
611 *californicus*. *Molecular Ecology Resources*. 15, 868-879.

612

613 Burton, R.S., Feldman, M.W., 1981. Population genetics of *Tigriopus californicus*. II.
614 Differentiation among neighboring populations. *Evolution*. 35, 1192-1205.

615

616 Burton, R., Lee, B., 1994. Nuclear and mitochondrial gene genealogies and allozyme
617 polymorphism across a major phylogeographic break in the copepod *Tigriopus*
618 *californicus*. *Proc. Of The National Academy Of Sciences*. 91, 5197-5201.

619

620 Edmands, S., Deimler, J.K., 2004. Local adaptation, intrinsic coadaptation and the
621 effects of environmental stress on interpopulation hybrids in the copepod *Tigriopus*
622 *californicus*. *J. of Experimental Marine Biology and Ecology*. 303, 183-196.

623

624 El-Khoury, R., Dufour, E., Rak, M., Ramanantsoa, N., Grandchamp, N., Csaba, Z.,
625 Duvillie, B., Benit P., Gallego, J., Gressens, P., Sarkis, C., Jacobs, H.T., & Rustin, P.,
626 2013. Alternative oxidase expression in the mouse enables bypassing cytochrome c
627 oxidase blockade and limits mitochondrial ROS overproduction. *PLoS Genetics*. 9,
628 e1003182.

629

630 El-Khoury, R., Kemppainen, K.K., Dufour, E. Szibor, M., Jacobs, H.T., Rustin, P., 2014.
631 Engineering the alternative oxidase gene to better understand and counteract
632 mitochondrial defects: state of the art and perspectives. *Br. J. of Pharmacology*. 171,
633 2243-2249.

634 Fernandez-Ayala, D.J., Sanz, A., Vartiainen, S., Kemppainen, K.K., Babusiak, M.,
635 Mustalahti, E., Costa, R., Tuomela, T., Zeviani, M., Chung, J., O'Dell, K.M.C., Rustin, P.,
636 Jacobs, H.T., 2009. Expression of the *Ciona intestinalis* Alternative Oxidase (AOX) in
637 *Drosophila* Complements Defects in Mitochondrial Oxidative Phosphorylation. *Cell*
638 *Metabolism*. 9, 449-460.
639
640 Genova, M.L., 2014. Electron Transport in the Mitochondrial Respiratory Chain. In:
641 Hohmann-Marriott, M.F. (Ed.), *The Structural Basis of Biological Energy Generation*,
642 vol. 39. *Advances in Photosynthesis and Respiration*. Springer, Switzerland, pp. 401-
643 417.
644
645 Hakkaart, G.A., Dassa, E.P., Jacobs, H.T., Rustin, P., 2006. Allotopic expression of a
646 mitochondrial alternative oxidase confers cyanide resistance to human cell respiration.
647 *EMBO Reports*. 7, 341-345.
648
649 Hong, B.C., Shurin, J.B., 2015. Latitudinal variation in the response of tidepool
650 copepods to mean and daily range in temperature. *Ecology*. 96, 2348-2359.
651
652 Kemppainen, K.K., Rinne, J., Sriram, A., Lakanmaa, M., Zeb, A., Tuomela, T.,
653 Popplestone, A., Singh, S., Sanz, A., Rustin, P., Jacobs, H.T., 2014. Expression of
654 alternative oxidase in *Drosophila* ameliorates diverse phenotypes due to cytochrome
655 oxidase deficiency. *Human Molecular Genetics*. 23, 2078-2093.
656
657 Lauritano, C., Procaccini, G., Ianora, A., 2012. Gene expression patterns and stress
658 response in marine copepods. *Marine Environ. Res.* 76, 22-31.
659
660 Liu M, Guo X., 2017. A novel and stress adaptive alternative oxidase derived from
661 alternative splicing of duplicated exon in oyster *Crassostrea virginica*. *Science Rep.* 7,
662 10785.
663
664 May, B., Young, L., Moore, A.L., 2017. Structural insights into the alternative oxidases:
665 are all oxidases made equal? *Biochem. Soc. Trans.* 45, 731-740.
666
667 McDonald, A.E., Vanlerberghe, G.C., 2004. Branched mitochondrial electron transport
668 in the Animalia: presence of alternative oxidase in several animal phyla. *IUBMB life*. 56,
669 333-341.
670
671 McDonald, A.E., 2008. Alternative oxidase: an inter-kingdom perspective on the function
672 and regulation of this broadly distributed 'cyanide-resistant' terminal oxidase. *Functional*
673 *Plant Biology*. 35, 535-552.
674
675 McDonald, A.E., 2009. Alternative oxidase: what information can protein sequence
676 comparisons give us?. *Physiologia plantarum*. 137, 328-341.
677

678 McDonald, A.E., Vanlerberghe, G.C., Staples, J.F., 2009. Alternative oxidase in
679 animals: unique characteristics and taxonomic distribution. *J. of Experimental Biology*.
680 212, 2627-2634.

681
682 McDonald, A.E., Gospodaryov, D.V., 2018. Alternative NAD(P)H dehydrogenase and
683 alternative oxidase: Proposed physiological roles in animals. *Mitochondrion*. In press.

684
685 Neimanis, K., Staples, J.F., Hüner, N.P.A., McDonald, A.E., 2013. Identification,
686 expression, and taxonomic distribution of alternative oxidases in non-angiosperm
687 plants. *Gene*. 526, 275-286.

688
689 Raisuddin, S., Kwok, K.W., Leung, K.M., Schlenk, D., Lee, J.S., 2007. The copepod
690 *Tigriopus*: A promising marine model organism for ecotoxicology and environmental
691 genomics. *Aquatic Toxicol.* 83, 161-173.

692
693 Robertson, A., Schaltz, K., Neimanis, K., Staples, J.F., McDonald, A.E., 2016.
694 Heterologous expression of the *Crassostrea gigas* (Pacific oyster) alternative oxidase in
695 the yeast *Saccharomyces cerevisiae*. *J. Bioenerg. Biomembr.* 48, 509-520.

696
697 Rodriguez-Armenta, C., Uribe-Carvajal, S., Rosas-Lemus, M., Chiquete-Felix, N.,
698 Huerta-Ocampo, J.A., Muhlia-Almazan, A., 2018. Alternative mitochondrial respiratory
699 chains from two crustaceans: *Artemia franciscana* nauplii and the white shrimp,
700 *Litopenaeus vannamei*. *J. Bioenerg. Biomembr.* 50, 143-152.

701
702 Rogov, A.G., Sukhanova, E.I., Uralskaya, L.A., Aliverdieva, D.A., Zvyagilskaya, R.A.,
703 2014. Alternative oxidase: distribution, induction, properties, structure, regulation, and
704 functions. *Biochem.* 79, 1615-1634.

705
706 Schertl, P., Braun, H-P., 2014. Respiratory electron transfer pathways in plant
707 mitochondria. *Frontiers in Plant Science*. 5, 163.

708
709 Sussarellu, R., Fabioux, C. Sanchez, M.C., Le Goïc, N., Lambert, C., Soudant, P.,
710 Moraga, D., 2012. Molecular and cellular response to short-term oxygen variations in
711 the Pacific oyster *Crassostrea gigas*. *J. of Experimental Marine Biology and Ecology*.
712 412, 87-95.

713
714 Van Aken, O., Giraud, E., Clifton, R., Whelan, J., 2009. Alternative oxidase: a target and
715 regulator of stress responses. *Physiologia plantarum*. 137, 354-361.

716
717 Vanlerberghe, G.C., 2013. Alternative oxidase: a mitochondrial respiratory pathway to
718 maintain metabolic and signaling homeostasis during abiotic and biotic stress in plants.
719 *International J. of Molecular Sciences*. 14, 6805-6847.

720
721 Yusseppone, M.S., Rocchetta, I., Sabatini, S.E., Luquet, C.M., del Carmen Rios de
722 Molina, M., Held, C., Abele, D., 2018. Inducing the alternative oxidase forms part of the
723 molecular strategy of anoxia survival in freshwater bivalves. *Frontiers in Physiol.* 9, 100.

- 724 Zalutskaya, Z., Lapina, T., Ermilova, E. 2015. The *Chlamydomonas reinhardtii*
725 alternative oxidase 1 is regulated by heat stress. *Plant Physiol. Biochem.* 97, 229-234.
726
- 727 Zhang, H., Finiguerra, M., Dam, H. G., Huang, Y., Xu, D., Liu, G., Lin, S., 2013. An
728 improved method for achieving high-quality RNA for copepod transcriptomic studies. *J.*
729 *of Experimental Marine Biology and Ecology.* 446, 57-66.